

THE THERMAL INACTIVATION OF PSYCHROPHILIC BACTERIA  
IN MILK

by

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ABSTRACT

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To investigate the occurrence of thermophilic psychrophilic bacteria in milk after pasteurization by the High Temperature, Short Time (HTST) method, duplicate samples were removed in screw cap test-tubes after the holding section from four commercial HTST units. Standard Plate Counts (SPC) incubated 2 days at 32°C. and psychrophilic plate counts incubated 10 - 15 days at 4 - 5°C. were made on one duplicate. The other duplicate was kept un-opened at 4 - 5°C. for 10 days at which time SPC and psychrophilic plate counts were made on it. SPC and psychrophilic plate counts were also made on the raw milk.

When no significant pattern of psychrophilic survivors was found in pasteurized milk held for 10 days, a limited laboratory experiment was initiated. Duplicate one millilitre amounts of milk containing high concentrations of three psychrophilic strains of Pseudomonas fluorescens were given an HTST heat treatment in sealed glass containers made from glass tubing of 3 mm. bore and 1 mm. wall thickness. An innovation was the use of a copper-constantan thermocouple sealed into some tubes of milk and attached to a recording potentiometer

to show both the time and temperature of the entire heating and cooling process.

Psychrophilic plate counts were made immediately after pasteurization on one glass tube duplicate at 4 - 5°C. for 10 - 15 days. The other glass tube duplicate was held un-opened at 4 - 5°C. for 10 days and then used for a second psychrophilic plate count.

No survivors were found immediately after pasteurization among the three strains of Ps. fluorescens. One strain showed survivors at a low level after the 10 days storage.

It is possible that some thermophilic strains of psychrophilic bacteria do exist. However, when the data of the first experiment is considered also, it seems unlikely that a thermophilic psychrophile will occur in normal raw milk supplies in sufficient numbers to survive HTST pasteurization. Hence, it is concluded that psychrophilic bacteria in milk are more likely the result of post-pasteurization contamination.

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INTRODUCTION



## INTRODUCTION

Psychrophilic bacteria are a significant obstacle to keeping milk fresh under refrigeration. It has been assumed, generally, that these organisms gain entrance to the milk after pasteurization. If it can be shown that these bacteria also survive pasteurization, then psychrophilic bacteria deserve greater attention in assessing the quality of raw milk. Using four psychrophilic strains of Pseudomonas fluorescens and the Modified Small Plate Culture Method for samples exposed to High Temperature Short Time (HTST) type pasteurization as outlined in the 10th Edition of Standard Methods (1), Macaulay et al (56) found highly significant numbers of surviving test organisms in 5 ml. of milk held for ten days at refrigerated storage temperatures.

Prompted by Macaulay's findings, the first experiment was undertaken to detect and enumerate the presence in milk of viable psychrophilic bacteria immediately after an HTST pasteurization. The four commercial HTST pasteurizers used plate type heat exchangers to heat milk to a minimum of 161.5°F. and approved holding tubes to maintain this temperature for 16 - 20 seconds.

The second experiment grew out of the failure to detect any significant numbers of bacteria capable of growing at 4°C. in milk collected from the end of the holding tube of the four HTST pasteurizer units. Because the experiments of Macaulay had found survivors of Pseudomonas species in milk given a heat treatment in the range 162.5 - 163.9°F. (72.7 - 73.3°C.) for 16 - 20 seconds estimated time of exposure, it was decided to repeat his experiment in part using slightly different methods and materials for exposing the bacteria to the same time/temperature treatment.

The major innovation of this experiment is the adaptation of a semi-micro sealed glass tube, total immersion technique to an HTST treatment, and the control of the time and temperature of heating and cooling with the use of a thermocouple and recording potentiometer.

Because the numbers of psychrophiles surviving in pasteurized milk, if any, are few, a 10 day incubation period at 4°C. was decided on to give any survivors a chance to grow. It was considered that if the lag period of survivors was any greater, then for all practical purposes, they were inactive and not a keeping quality problem. To show the extent

of deviation in viable bacterial counts obtained from freshly pasteurized milk as compared to counts from the same milk held at 4°C. for 10 days, duplicate samples were used. One was to be counted as soon as possible, the other in ten days' time, with Standard Plate counts (SPC) and Psychrophilic Counts being run each time. This same pattern was followed in both experiments.

LITERATURE REVIEW

## LITERATURE REVIEW

Various definitions have been advanced for the term "psychrophile" or "cold-loving" as applied to bacteria. Fabian (29) classifies cryophilic, mesophilic, and thermophilic bacteria according to their minimum, optimum, and maximum growth temperatures. Cryophilic bacteria are defined as having a minimum of 32°F., an optimum of 59°F. and a maximum of 86°F. Mesophiles are defined as having a minimum of 59°F., an optimum of 98.6°F. and a maximum of 113°F. Thermophiles have a minimum of 113°F., an optimum of 131°F and a maximum of 158°F.

Kennedy and Weiser (50) define psychrophiles as having an "optimum temperature range" of 5 - 25°C. They cite an optimum of 25 - 45°C. for mesophiles.

Olsen et al (65) cites 35 - 45°F. as the psychrophile range. Thomas (92) defines them as able to grow at less than 5°C. and unable to survive 63°C. for 30 minutes.

Oginsky and Umbreit (62) classify psychrophile's optimum temperature as 12 - 18°C. with an approximate minimum of 0°C. They place mesophile's optimum range as 25 - 37°C. or 37 - 50°C. with an approximate minimum of 15°C. Thermophiles have an

optimum of 55 - 60°C., a minimum of 40°C., and a maximum of 75°C. Ingraham and Stokes (46) define psychrophilic bacteria as those that grow well at 0°C. within two weeks' time.

Prouty (74) uses the adjective "facultative" to describe bacteria of mesophile optimum growth temperature which are capable of significant growth under psychrophilic conditions. Roth (82) has demonstrated with the genus Arthrobacter a continuous progression from psychrophile to mesophile of seven species' response to growth temperatures. Regardless of their definition, Elliker (23) says that these psychrophiles are the Dairy Industry's biggest problem.

#### LOW TEMPERATURE KEEPING QUALITY

It is generally recognized that keeping quality should be consistent. If a housewife uses a half pint of whipping cream over two weeks, she expects the next to keep as long. When all the dairy plants in a given area have the same keeping quality in their products, none of them is likely to suffer by comparison. Thus there is no incentive for improvement, even if the keeping quality of all of them is mediocre. As pointed out by Olsen (65), Boyd (8), Ford (30), Speck (87), and Elliker (23),

the trend to increased marketing areas, rising labour costs, the expense of automation and the investment in large scale equipment coupled with the handling of raw milk in bulk quantities and the transport of pasteurized milk over long distances all makes improved quality imperative.

In 1959, Ford (30 ) stated that four days was a common length of time between processing and consumption. With wider areas being served and larger size containers such as three quart cartons, the length of time between processing and consumption is increasing. Inventory control can be improved and the waste of scrapping older packages decreased where management has confidence in their product's keeping quality.

#### HIGH TEMPERATURE - SHORT TIME PASTEURIZATION

The heat treatment of milk known as Pasteurization was first used about 1890. From its first introduction, the equivalence of exposures using higher temperatures for shorter times to lower temperatures for longer times was recognized. Because the lethal length of time at higher temperatures became a matter of seconds and because of a tendency to disperse fat globules so that no cream layer formed, a lower temperature for a longer, more easily

controlled, time was chosen.

Since it was the most heat resistant pathogen likely in milk, the destruction of M. tuberculosis was adopted as the standard by which equipment and heat treatments were judged. The time and temperature exposure first agreed to was 143°F. for 30 minutes. This contained a ten minute safety margin over the time required to kill M. tuberculosis and did not affect cream line formation. Much later, when the importance of cream line formation had declined, Enright ( 24 ) showed that Rickettsia burneti, the cause of Q fever, would survive the Holder method (143°F. for 30 minutes). Raising the Holder method treatment to 145°F. for 30 minutes resulted in complete destruction of R. burneti. The same author ( 25 ) found increased temperature and time necessary where ice cream mix, chocolate milk and cream were processed.

Holmquist ( 45 ) relates that the first licensed commercial High Temperature - Short Time unit had two electrodes immersed in the milk for heating. With fixed electrical input, control was by flow rate from a pump with an automatic stop. The use of hot water heating devices worked as well when it was realized that ".....the degree of heat and the holding time rather than the method of applying the



heat are the important factors" ( 45, p. 14).

Weber (102) states that the initial time-temperature treatment agreed upon for HTST was 160°F. for 15 seconds. The introduction of the flow diversion valve, a safety device to prevent sub-heated milk from moving beyond the heating and holding sections of the apparatus, necessitated increasing the length of holding time one second to 16 seconds. This added second gave time for the valve to operate. Kreuger ( 52) states that the temperature was raised one degree (1°F.) as an added precaution. In checking the original exposure, Workman (103) tested 17 strains of human and bovine tubercle bacilli, 74 strains of Brucella abortus, 218 strains of human and 186 strains of Bovine streptococci at 160°F. plus or minus 5°F. for 15 seconds in a laboratory pasteurization set up. In no case were any survivors found among all these micro-organisms.

In comparing plate counts of milk pasteurized by the Holder method (143°F. for 30 minutes) and by the HTST method, occasional high counts were found in the HTST pasteurized milk. Parfitt ( 69), Prucha and Parfitt (75 ), Fabian ( 29), and Kreuger ( 52)(53) found that these were thermoduric organisms that originated on the farm and that they could be controlled

by improved sanitation, particularly in the care of rubber milking machine parts.

Hileman (42) (43) in 1941 found a difference in kinds of microflora surviving when comparing Holder and HTST methods. Increased numbers of alkali producers found by litmus milk reactions were noted in plant HTST milk. From commercial experience over an eighteen month period, he dismissed them as being of no importance as judged by complaints about undesirable flavours and odours. It is well to bear in mind that delivery schedules were then more frequent.

KEEPING QUALITY RELATED TO TIME AND TEMPERATURE OF STORAGE, BACTERIAL GROWTH AND FLAVOUR DEVELOPMENT

Numerous studies have been made in the past of keeping quality or the length of time before bacterial growth causes off-flavours, odours, and other changes in milk that has been pasteurized. Olsen (65) lists flavours as "unclean, putrid, fruity, unclean sour flavour". Further defects are thickening, ropiness, stringiness, and a slight green or yellow colouration. Erdman and Thornton (26)(27) reported bitter flavours in milk held 15 days at 4.5°C. Kastli and Binz (48) found oxidized flavours in samples stored at 2 - 3°C. after 2 - 3 days' storage. Atherton et al (5) found off-flavour development

related to a decrease in the protein stability and that "stale" flavours were the most usual. Boyd ( 9 ) reported no oxidized flavours among the off-flavours due to growth of psychrophiles in milk.

Sherman ( 84 ), in studies using 0°C. storage, kept good quality pasteurized milk for periods of eight to twelve weeks. He further found that good quality raw milk, capable of keeping up to four weeks, could decrease the keeping quality of the pasteurized milk when injected in minute quantities into the latter. He found no spore formers growing, and blamed nearly all growth on Pseudomonas species.

In a more recent study of retail pasteurized milk in Lexington, Kentucky, Randolph et al (78) found that the average time for milk to keep a satisfactory flavour was 7.7 days. Further, it was found that after 10 days, 58% of samples were off-flavour, and after 14 days, 74% were off-flavour.

Boyd et al ( 9 ) found that keeping quality shown by flavour scores extended beyond the time that it took the psychrophile count to reach 50,000/ml. He found lowering the temperature of storage from 40°F. to 33°F. increased keeping quality time 11 to 14 days. Keeping quality at 33°F. was approximately the same using a 50,000/ml. standard and a flavour

score of 37. Broitman, Mallmann and Trout (10) found off-flavours in pasteurized milk with counts ranging from  $8 \times 10^6$  to  $1 \times 10^8$  per millilitre.

Other studies, involving the keeping quality of commercially pasteurized milk, are those of Day and Doan (20), Nelson and Baker (61), Rogick and Burgwald (81), and Burgwald and Josephson (11). They all indicate the association of off-flavour development and the growth of psychrophilic bacteria.

#### TYPES OF BACTERIA CAUSING STORAGE PROBLEMS

Elliker (23) lists the following genera as being the most frequently encountered in keeping quality problems of pasteurized milk and cream: Pseudomonas, Achromobacter, Chromobacterium, Alcaligenes, Proteus, Escherichia, and Aerobacter. The genus found most frequently to be the cause of both high counts and flavour defects is Pseudomonas. Punch et al (76) found that 24 pure cultures of Pseudomonas gave off-flavours in HTST pasteurized milk with counts ranging from 5.2 - 200 million. Using gas chromatography, Mickelson et al (59) found the flavour produced by a Pseudomonas fluorescens to consist of four different components.

From commercially pasteurized milks, Pseudomonas spp. have been isolated by the following:

Lawton and Nelson (54) who found 7 strains; Hempler (41) after storage at 7°C. found Pseudomonas along with Achromobacter and Alcaligenes; Baumann and Reinbold (6) found Pseudomonas in six out of ten samples classified. Galesloot (34) states that they are the principal cause of milk spoilage under refrigeration; while Parker et al (71) state that the genus is the most important to the whole dairy industry.

From raw milks, numerous investigators have isolated psychrophiles of the Pseudomonas genus. Thomas, Hobson, and Bird (93) found that of 102 psychrophile isolates, 99 were gram negative rods and of these, 35 out of the original 102 were Ps. fluorescens. Andrey and Frazier (3) found that Pseudomonas spp. were the second most frequent isolate. Morris (60) found that Pseudomonas growth materially contributed to the failure of the methylene blue test by certain producers, and traced it to the farm water supply. In an extensive study of farm bulk tanks, Marth and Frazier (57) tested various genera isolated from raw bulk tanks for growth ability at 38°F. when re-inoculated into raw milk. Pseudomonas spp. grew more rapidly than Achromobacter, Alcaligenes, and Aerobacter toward the end of the four day test period. Also, a difference in the ability of some cultures to grow

better in raw than in either pasteurized or sterilized milk was observed. Erdman and Thornton (27) identified 190 cultures out of 722 psychrophile isolates from raw milk and cream. Of these, Pseudomonas spp. accounted for 16.7% and also for over half of the Gram negative rods of which 62% exhibited fluorescence.

Defects in flavour and other qualities of both cottage cheese and butter have been caused by psychrophilic organisms. Large proportions of these have been Pseudomonas spp. traced to the water supply. With water and butter samples from Wisconsin and Minnesota creameries, Jezeski and Macy (47) found that twenty-eight out of forty-one cultures isolated were of the genus Pseudomonas and that a majority of these produced a green fluorescence in culture media. Druce and Thomas (21) studied farm butter made using un-chlorinated and contaminated water. Pseudomonas putrefasciens and fluorescent Pseudomonas spp. were found among the large number of psychrophilic bacteria counted. In his investigation of an intermittent surface taint of butter, Schutt (83) found that Pseudomonas fluorescens was the most important organism. Corley and Hammer (16) traced the spoilage of butter to high counts of Ps. fluorescens and Ps. putrefasciens in creamery water supplies.

Parker et al ( 70 ) and Davis and Babel ( 19 ) investigated a slimy curd defect of cottage cheese. Parker isolated a Pseudomonas viscosa that caused a yellowish or brownish slime and extensive proteolysis. The same author found that Pseudomonas fragi would cause a fruity odour. Davis and Babel found several Pseudomonas spp. that would cause a slimy curd defect. Bonner and Harmon ( 7 ) found Pseudomonas desmolyticum, Ps. fragi, Ps. fluorescens, and Ps. tralucida among samples of spoiled cottage cheese. In their survey of cottage cheese quality, Martin, Foltz, and Rutz ( 58 ) found that 44% of all samples examined had Pseudomonas species present. Like Parker above, they found that Pseudomonas viscosa gave a yellowish brown slime along with fruity, rancid, bitter, and flat flavour defects.

#### CHARACTERISTICS OF PSEUDOMONAS SPECIES AND OTHER PSYCHROPHILES

##### Temperature of Growth

Different Pseudomonas species have been found having a very wide range of growth temperatures. Sulzbacher ( 91 ) found isolates from frozen meat, apparently Pseudomonas spp., that grew on agar slants at  $-6^{\circ}\text{C}$ . ( $21^{\circ}\text{F}$ .). They produced a yellowish green pigment and alkaline reduction in milk.

Straka and Stokes ( 89 ) found isolates from Antarctica that grew on Trypticase Soy Agar at  $-7^{\circ}\text{C}$ . Their maximum growth temperature was  $35^{\circ}\text{C}$ . for 28 out of 31 strains. Only two grew at  $37^{\circ}\text{C}$ .

Olsen and Jezeski ( 63 ) grew a psychrophilic strain of Pseudomonas fluorescens in both aerated and stationary cultures at 4, 10, 15, 20, 25 and  $32^{\circ}\text{C}$ . They found that aeration produced more rapid growth at low temperatures. In their study of farm bulk tank flora, Marth and Frazier ( 57 ) isolated six species that grew well at  $38^{\circ}\text{F}$ . Their Pseudomonas species' growth rates in milk at bulk tank temperatures were such that initial counts in the range of 50,000 - 115,000 could reach 200,000/ml. in 24 hours. An initial count of 30,000 - 70,000/ml. would reach 200,000/ml. after 48 hours under the same growth conditions.

Castell and McDermott ( 12 ) investigated the growth characteristics of organisms found in unchlorinated tap water from a 220 foot deep drilled well. They found many psychrophilic types on plates incubated at  $3^{\circ}\text{C}$ . that were active multipliers in the water. After one week of growth in the well water, 100% resembled Pseudomonas fluorescens. Of 60 species chosen for study the most dominant and important were



gram negative, alkaline forming, oxidase positive, proteolytic and lipolytic organisms with a relatively low temperature range.

Psychrophiles able to grow in the absence of oxygen are reported by Upadhyay and Stokes ( 97) Under both aerobic and anaerobic conditions, decreasing incubation temperatures lengthened lag, exponential, stationary, and death phases. Elimination of oxygen increased the lag period, allowed the cells to survive much longer at low temperatures (but accelerated their death at 20°C. and higher), and finally, reduced the extent of growth. Maximal cell populations were obtained aerobically at 5°C., and anaerobically at 25°C. No species study was made. These findings contrast to the cold-loving, strictly aerobic organisms isolated by Straka and Stokes (89 ) from Antarctica.

#### Nutritional Requirements of Pseudomonas and other Psychrophiles

The well water of Castell and McDermott ( 12) on chemical analysis showed these ions present:  $\text{NH}_4^+$ ,  $\text{Cl}^-$ ,  $\text{Ca}^{++}$ , and  $\text{Mg}^+$ . Trace amounts of carbon were detected when 100 ml. of the well water was evaporated to dryness. Carbohydrate fermenters, gram positive cocci, and spore forming bacilli were almost

entirely absent. The organisms were able to use salts of organic acids.

In discussion of nutritional requirements of psychrophiles causing spoilage in cottage cheese, Harmon (38) says that Pseudomonas and Alcaligenes genera are not fastidious. However, Straka and Stokes (88) showed that with lower temperatures, Pseudomonas fluorescens required a more complex media, Trypticase Soy (contains trypticase, a pancreatic digest of casein, and phytone, a papaic digest of soya-bean meal). A contrasting simple glucose-salts media supported no growth. These authors suggest that the more complex media overcomes "injury" caused by the low temperature. That heating above their normal temperature range also injures is suggested by the findings of Heather and Vanderzant (40) who showed that Ps. fluorescens held at 55°C. showed most survivors of longer heating times when grown on complex media. These same authors (39), (40) report a greater lag phase of heated cultures of Ps. fluorescens, Ps. fragi, and Ps. putrefasciens.

Olsen and Jezeski (63) compared glucose, monosodium citrate, and casamino acid as primary carbon sources for a psychrophilic strain of Pseudomonas fluorescens. The most rapid growth at 4°C.

was with the casamino acids. The growth stimulation of the casamino acids was due to two possible situations. They say that either the casamino acids provided substrate material as precursors needed in cell synthesis or build-up, or they provided substrates suited to catabolic or breaking down action by the cell at low growth temperature.

Because high psychrophile counts have been noted in the pasteurized product from plants whose raw supply had a high count, investigation as to any influence of growth prior to pasteurization on subsequent growth was made. Overcast and Adams (67) found that excessive psychrophile growth in raw milk stimulated Brevibacterium lipolyticum and a psychrophile isolate, inhibited Pseudomonas fragi after 1, 2, and 4 days growth, and had no stimulation for Ps. fluorescens when these were grown in laboratory pasteurized milk. Their failure to prove the subsequent stimulation hypothesis is supported by earlier work of Sinclair and Stokes (85). Using Ps. fluorescens in a chemically defined liquid media containing 1% glucose and 0.1%  $(\text{NH}_4)_2\text{SO}_4$ , they found that the organism grew at 30°C. until it used up both compounds. If the energy and nitrogen sources were added back to the filtrate of the growth media, the

Ps. fluorescens reinoculated would reach maximal levels as in the original culture. After several repetitions, mineral deficiencies resulted. When replaced, growth occurred again. They concluded that there was no evidence that auto-inhibitory substances were made or that physical crowding limits growth. In a similar experiment with Escherichia coli, organic acids accumulated and inhibited growth.

#### Enzyme Activity of Pseudomonads and Psychrophiles

Lipolytic and proteolytic activity were reported in the isolates of Castell and McDermott (12). Overcast and Skean (68) found isolates of Pseudomonas, Achromobacter, Alcaligenes, and Flavobacterium genera all capable of hydrolyzing butterfat at 4°C. Thomas, Hobson, and Bird (93) found that most of their gram negative isolates hydrolyzed tributyrin but that only one-third hydrolyzed tri-olein. Druce and Thomas (21) found that farm butter with a high yeast and mould count and large numbers of psychrophile and mesophile bacteria became rancid in two or three days. Green and Jezeski (37) found two isolates classified as Pseudomonas, one of which was distinctively lipolytic, while the other was proteolytic. Parker et al (70) observed extensive proteolysis by a Pseudomonas viscosa.

In making detailed proteolytic studies of their 102 psychrophile isolates of which 99 were gram negative rods, Thomas, Hobson, and Bird (93) found that a "high proportion" liquefied gelatin, a "majority" showed slow proteolysis in litmus milk at 22°C., and slightly more than half showed caseolytic activity on 30% milk agar. Vanderzant and Moore (100) suggest that proteolytic activity of psychrophiles may vary with temperature.

Further studies on the influence of temperature on the enzyme activities of psychrophiles were carried on by Upadhyay and Stokes (98), (99). They found that a formic hydrogenlyase, and a hydrogenase with its enzyme forming system were different in temperatures of optimum activity and inactivation. The psychrophile enzyme source was a rod shaped, gram negative bacterium that grew well in the range of 0 - 35°C. When the psychrophile hydrogenase (99) was held at 60°C., there was 50% destruction of it after 2 hours as compared to only 25% destruction of a mesophilic hydrogenase under the same conditions.

#### Heat and Pasteurization Studies of Psychrophiles

Heat studies on psychrophilic bacteria have sought either to find the thermal death time of an organism, or to determine its survival of pasteurization.

Pasteurization in the dairy industry may legally describe two time/temperature relationships. The first is called Low Temperature, Long Time (or LTLT) pasteurization; the second, High Temperature, Short Time (or HTST) pasteurization. The Low Temperature, Long Time pasteurization, also known as the Holder method, raises every particle of the milk to a temperature of 62 - 63.5°C. for 30 - 35 minutes. The High Temperature, Short Time treatment is at a minimum of 71.6°C. for 16 seconds. Under laboratory conditions and with full size commercial pasteurizing units, the LTLT method is usually a batch process while HTST is continuous.

Because temperature and time are easily controlled, the LTLT treatment has been used the most to study the thermoduric properties of psychrophilic bacteria in the laboratory. When comparing different studies, the initial number of organisms with their previous growth and temperature history, and the heating, cooling and enumeration should be known. With small numbers and long plate incubation times at low temperatures, otherwise inconsequential contamination grows in importance.

#### Influence of Initial Numbers on Survivors

Using glass tubes as a container in which

various concentrations of Escherichia coli were heated, Craige (17) found survivors to a treatment of  $62 \pm 0.1^{\circ}\text{C}$ . for 30 minutes. Various strains survived at concentrations ranging from  $\log_{10} 4.2$  -  $\log_{10} 12$  organisms per millilitre. The enumeration technique involved pouring the inoculated pasteurized milk from a glass heating tube into lactose broth fermentation tubes. Both Prouty (73) and Stumbo (90) point out the relation borne by numbers of bacteria to the time-temperature treatment needed for their destruction. When number of survivors is plotted logarithmically and time is plotted linearly on semi-logarithmic paper, the numbers surviving for successive units of time follows a straight line. This gives a rate of destruction curve.

The use of thermal death time curves has been made by Kaufman and Andrews (49), and Chaudhary, Tuckey and Winter (13) to show that psychrophiles are unlikely to survive pasteurization. Kaufman and Andrews used two psychrophile organisms, one of them a Pseudomonas viscosa - 3, at three different temperatures below LTLT temperature in order to determine their thermal death curves. (Of 66 isolates from milk and water, Kaufman and Andrews found none that survived LTLT pasteurization in the laboratory).

Olsen, Macy, and Halvorsen (64) state that plotting "z" curves in the above manner explained differences in survivors of the two pasteurization heat treatments. However, in a recent discussion of "Ultra-High" temperature and time heat treatments for the pasteurization of milk, Robinson (79) cautions that extrapolation of thermal death time data gained at lower temperatures is not justified.

Many investigators have used an LTLT treatment under laboratory conditions to determine thermotolerant or heat resistant properties of psychrophilic bacteria: (7), (47), (27), (50), (94), (95), (5), (70), (49), (67).

When the risk of extrapolation to the higher temperature used for HTST is borne in mind, the information from laboratory studies in regard to Pseudomonas species in particular and psychrophiles in general is vague. Jezeski and Macy (47) found that 6 out of 41 psychrophile isolates survived 65.5°C. for 30 minutes. Twenty-eight of the forty-one were Pseudomonas species. Nowhere do they state that any of the surviving six were Pseudomonas spp. Erdman and Thornton's (27) psychrophile survivors of 142°F. (61°C.) were gram negative non-fluorescing cocci. Kennedy and Weiser (50) had two of twelve pure cultures that



showed only 5% and 24% reduction of their numbers at 145°F. for 30 minutes. These had been isolated at 10°C.

Most of the above studies used some sort of tube immersed in a water bath which experience has shown is well suited to LTLT studies. The necessity to study milk and its flora at higher temperatures has resulted in a variety of other means and equipment. To study creaming and phosphatase tests for pasteurization with HTST in the early 1940's, Holland and Dahlberg ( 44 ) used a thin walled copper container, 12 by 20 by 1 centimeter, that they immersed in water. Franklin ( 32 ) has developed a similar device quite recently that heats the milk between surfaces only 5/32 inch apart.

The use of liquid media into which large numbers of organisms could be injected and then retrieved at timed intervals from a constant temperature state are described by Gilcreas ( 35 ), Kaufman and Andrews ( 49 ), Chaudhary et al ( 13 ) and Foster ( 31 ) This method has been found suited to thermal death time studies ( 49 ), ( 13 ) where automatic devices to remove bacteria and media from the test chamber were devised ( 35 ). Enright ( 25 ) devised a more sophisticated apparatus where liquid media or milk was placed

in a 3 millimeter clearance between two concentric cylinders. The inner cylinder rotated inside the sealed outer cylinder giving constant and continuous agitation. A thermocouple and continuous recorder gave time and temperature curves of the heat treatment when the apparatus was heated by immersion in a water bath.

The use of commercial units to test for the survival of psychrophiles in HTST and LTLT treatments was made in "in plant" studies by Rogick and Burgwald (80 ), (81 ). In neither treatment could they find any bacteria in 4.1 millilitres of milk immediately after pasteurization with plates incubated at 4 - 7°C. for twelve days. However, after 7 days' storage in a refrigerator at 4 - 7°C., all their samples showed some psychrophile count (on plates again incubated at 4 - 7°C.). However, it should be noted that the counts made after 7 days' storage in a refrigerator, were made on the same samples used for the counts made immediately after pasteurization.

With initial numbers not reported, Olsen et al ( 65) found no survivors when sealed glass tubes of milk were totally immersed in a water bath for LTLT pasteurization. The psychrophile counts were all zero. These same authors withdrew milk aseptically from

between the flow diversion valve and the regeneration section of an HTST unit operated at 162°F. for 16 seconds holding time, stored it for seven days under refrigeration, and found psychrophile counts of three and zero.

In more recent trials comparing a High Temperature, Short Time treatment of 172°F. and a 16 second holding time to milk given an Ultra-High heat treatment, Glazier (36) found higher increases in counts with the HTST milk. Although the holding time used was 0.6 second and therefore not a directly comparable treatment to HTST, Evans, Lachman and Litsky (28) using 10°F. steps from 160 - 260°F. found significant psychrophile growth in milk processed in the 160 - 200°F. range and stored at 40°F.

#### Enumeration of Psychrophiles

Olsen et al (66) point out that bacteria that survive pasteurization show little growth at temperatures of 45°F. (7.2°C.) or lower. Consequently, psychrophilic counts on freshly pasteurized milk free of recontamination are nearly always negative if incubated at 40 - 45°F. for 7 - 10 days. This is supported by Thomas et al (94). Olsen et al (66) also point out the close relationship of plate counts made at 25, 32, and 35°C. and that 25°C. for 3 days will

include nearly all types of bacteria in the milk, including psychrophiles.

One method for detecting the presence of psychrophilic bacteria in pasteurized milk is the Moseley count ( 4 ) most recently favoured by Elliker (22 ). It involves storage at 45°F. (7.2°C.) for 5 days and then plating the milk from its retail container with plate incubation at either 25 or 32°C.

Attempts to selectively speed up the detection of psychrophiles have been made. Leesment (55) used preincubation at higher than usual temperatures, immediately after plating, before placing the plates in the more usual lower temperature surroundings. A surface plate technique of Punch and Olsen (77 ) reduced the time for colonies to appear by 2 - 3 days at 6°C. Day and Doan (20 ) and Csenge and Doan (18) developed the use of neotetrazolium under vacuum conditions as a dye reduction test capable of indicating psychrophile spoilage in advance of flavour defects. Collins (14 ) has used Violet Red Bile Agar as recommended by Elliker, to grow subsurface colonies in 48 hours at 25°C. of Pseudomonas fragi, Ps. viscosa, and Alcaligenes metalcaligenes. In a preliminary report, Freeman, Nanavati and Glen (33 ) have attempted to inhibit growth of gram positive organisms in order to

allow enumeration at a higher temperature of gram negative bacteria. They tested a number of chemicals with some measure of success.

Chemical Control of Psychrophiles and Pseudomonas

Castell and McDermot (12) reported that 3% salt (NaCl) and 1.5 parts per million (ppm) of chlorine would control the growth of the organisms found by them in well water. Collins (15) found that pH had a distinct bearing on the effectiveness of chlorine added to alkaline waters containing Pseudomonas species. Post and Krishnamurty (72) found considerable lysis of Pseudomonas fluorescens in the presence of sodium hexametaphosphate which is a common chelating or water softening agent for calcium and magnesium ions in hard water. Elliker (23) recommends wide-spread use and high (200 ppm) concentrations of chlorine to counteract the introduction of psychrophiles into dairy products under plant conditions.

EXPERIMENTAL METHOD

## EXPERIMENTAL METHOD

Experiment 1 - HTST Plant Pasteurization  
and Psychrophiles

The purpose of these experiments was to detect the presence of any psychophilic bacteria in milk immediately after heating in a plate type heat exchanger to a minimum of 161.5°F. and holding for 16 - 20 seconds in an approved holding tube.

Each raw storage tank was taken as the basic unit or lot of milk. Duplicate raw samples were taken from the end of the holding tube for each tank of milk at the beginning, middle, and end of the time required to process it. The samples were labeled and placed in wire racks in ice water. On return to the laboratory, SPC and psychrophile counts were made on one-half of each duplicate sample while each of the other halves were held at 4°C. for 10 days. At the end of the 10 days holding, SPC and psychrophile counts were repeated.

Sampling Technique

The sampling cock used at the end of the holding tube (as shown in Fig. 2) was a valve manufactured by the Aluminum Pressure Vessel (APV) firm of equipment manufacturers for the purpose of timing the holding period of their HTST units. The valve

threads into a normally plugged opening with a rubber O-ring gasket to seal it. The valve is equipped with a curved side arm that opens upward above the valve seat (in the opposite direction from the downward pointing delivery tube). This side arm allowed the delivery tube below the valve seat proper to be sanitized before each sample was taken, as shown in Fig. 2.

Prior to being taken to the dairy, the valve was sterilized in the autoclave at 248°F. for 20 minutes plus well wrapped in several layers of brown wrapping paper. At the dairy, the cock was fastened by the pasteurizing room employees to the end of the holding tube before the HTST unit was sanitized for the day's operation. Then the dairy's usual hypochlorite sanitizer was run through the cock for five to ten minutes. Before each sample was taken, the cock was sprayed with an iodine sanitizer all over its outside surface, and through the delivery tube and the side arm. Milk was allowed to run from the cock for about thirty seconds in order to rinse any sanitizer from the delivery tube before taking each split sample. The samples were collected in screw cap test-tubes which had been previously sterilized in the autoclave. Their capacity was approximately 25 millilitres and they were filled with about



20 millilitres of milk. Care was taken to see that the ice water level was higher than the level of the milk in the tubes.

#### Plating Procedure

Plate counts were run on both Plate Count Agar (Difco) and on Trypticase Soy Agar (B.B.L.) for the last three plant trials. Prior to this, counts were made on Plate Count Agar only. Each plate, on each media was made in duplicate. Poured plates were incubated for the SPC at 32°C. for 48 hours, and for 10 - 15 days for the Psychrophile Count at 4°C. Otherwise, the procedures followed were as outlined in Standard Methods ( 2 ).

#### Experiment 2 - HTST Laboratory Pasteurization of Psychrophiles

#### Test Organisms

Three Pseudomonas fluorescens strains as used by Macaulay et al ( 56) were used in this experiment:

#### Pseudomonas fluorescens

Strain - Spiers	Catalogue No. 3756
Strain - 85 Groombridge	Catalogue No. 9428
Strain - 25/8 ATCC 13525	Catalogue No. 10038

These three cultures were obtained through the courtesy of the National Collection of Type Cultures, Central Public Health Laboratory, Colindale,

London, NW 9. Obtained in lyophilized form, they were started in Trypticase Soy Broth, and then maintained on Trypticase Soy Agar Slants at 4 - 5°C. They were tested for growth at 37°C. and found to conform to Macaulay's information about them, that they were true psychrophiles unable to grow at 37°C.

For use with the glass tube heat treatment, these cultures were grown in Trypticase Soy Broth for 24 hours at 25°C. and then stored in the refrigerator. Transfers were made from these broth cultures with a .01 ml. loop into 250 ml. of Trypticase Soy Broth in 500 ml. capacity flasks. The flask contained a Teflon coated stirring magnet that was sterilized along with the media in the flask in the autoclave for 15 minutes at 15 psi. These flasks were stoppered with cotton plugs. Incubation was for 18 - 24 hours at 25°C. in a circulating, temperature controlled water bath. The magnetic stirrer was operated at a speed fast enough to form a slight vortex in the contents of the flask without the formation of foam.

To separate the organisms from the Trypticase Soy Broth, 40 ml. portions were pipetted into sterile plastic centrifuge tubes having a capacity of 50 ml. and centrifuged on a Clinical model centrifuge at 1560 g. The broth was decanted, and the cells

resuspended in 0.85% saline solution to the amount of 20 ml., and centrifuged down. The saline was then decanted. After a second saline washing using 20 ml., the cells were placed in two of the centrifuge tubes and spun down. The second saline washing was decanted, and a third amount of saline added to the amount of 3 - 5 ml. to give as high a concentration of cells as possible. This last step was performed using sterile Pasteur pipettes. This concentrate was placed in a clear glass test-tube, 17 mm. in diameter and a percent transmittance reading taken in a Lumetron Colorimeter at 620 mu. Then the suspension was examined in a Petroff-Hauser counting chamber using crystal violet (saturated alcoholic solution - 10 ml., placed in 100 ml. of distilled water) as recommended in the directions of the maker of the counting chamber (51)

Milk was obtained with different levels of test organisms as follows: using the counting chamber estimate of the number of organisms in the salt suspension, serial dilutions were made using 9 ml. quantities of homogenized milk which had been measured with a pipette into rubber stoppered test-tubes and sterilized for 5 minutes at 248°F. One millilitre of suspension was placed in the first tube of milk, mixed by inverting twenty-five times, and then one

millilitre of this was placed in the second 9 ml. of milk. This series of 1/10th dilutions continued so that the range of numbers of organisms in milk was from one-tenth the number in the original salt suspension to a concentration in the last tube of approximately 1000/ml. This series of suspensions in milk supplied the material to be sealed in glass tubes for the heat treatment. Psychrophile counts were made on those tubes used for pasteurizing material.

#### Sealed Glass Tube Pasteurization

The glass tubing used for heating and storing the milk was a standard type of soft flint glass with the following specifications:

Outside diameter	5 mm.	Softens - 700°C.
Inside diameter	3 mm.	Anneal - 520°C.
Wall thickness	1 mm.	

Coefficient of Expansion:  
 $92 \times 10^{-7}/1^{\circ}\text{C.}$  from 0 - 300°C.

This glass tubing came in 90 cm. lengths. These were cut to 23 cm. lengths. By filling a 900 mm. length of the tubing with water at room temperature, and measuring the amount delivered into a 10 ml. graduate, it was calculated that by filling the tube to a length of 14.1 cm., the amount delivered would be approximately 1 millilitre. This checked using the mathematical formula for the volume of a cylinder;

$$V = \pi r^2 D$$

where V = volume of cylinder  
 r = radius of cylinder  
 D = length of cylinder

Substituting: V = 1,000 cubic millimeters  
 r = 1.5 millimeters  
 D = unknown  
 $\pi = 3.14159$

$$1,000 = 3.14159 \times 1.5 \times 1.5 \times D$$

$$D = 141.4 \text{ mm.}$$

$$= 14.1 \text{ cm.}$$

The remainder of the length of the tube interior was used up partly by sealing and partly for an air cushion of 4 - 5 cm. that allowed for expansion and contraction of the milk during heating and cooling. Prior to being placed in pipette holders for sterilizing in the autoclave, one end of each tube was closed by holding and rotating it in an ordinary gas (Bunsen) flame until a solid glass plug formed. The milk suspensions were placed in the tubes with a sterile Pasteur pipette and filled from the sealed end to a distance of 14 cm. The milk was then sealed in the tube using the gas flame. When the hottest possible flame was used, the tube could be sealed without any heat being conducted to the milk. The operator could hold the tube at or beyond the level of the milk during the sealing without his bare fingers feeling any heat.

To hold the tubes in the hot water bath and then in the cooling ice bath, a plated wire test-tube

holder was adapted so that it would hold the tubes in a horizontal position. This was important in order to insure that all five tubes being heated came in contact with the heating water and the cooling ice water at the same instant. The test-tube rack was modified by using four zig-zag wires fastened to the middle platform. The tubes of milk were held in place by the spring of the wire and spaced approximately  $1/8$  inch apart. Two of the wires were used at either end of each set of tubes. With this arrangement, the five tubes could be immersed at a time. One of these tubes was a probe tube containing a thermocouple and also filled with milk. Thus all five tubes were close enough together that they would be heated and cooled the same amount.

The tubes were identified by the number of their position in the rack and so labelled during their 10 day storage.

Temperatures of cooling baths, heating, and incubation were measured using a mercury bulb thermometer. The measurement of the temperature of the milk in the glass tubes as well as the duration of the High Temperature, Short Time pasteurization exposure was accomplished with the use of a thermocouple and recording potentiometer. The copper-constantan

junction (the thermocouple) was welded on an impulse spot welder and so positioned in the middle of the 3 mm. diameter bore of the glass tubing that the wires leading to it did not touch the side for at least 1 cm. Centering the thermocouple was accomplished by bending the wires. This position could be checked by rotating the tube held up to the light. If it was off center, an eccentric wobble could be observed. The lead wires were sealed into the tubes at 2 cm. from the end in such a way that the sealing gas flame did not impinge directly on the wires at the point of sealing. The glass tube with the wires inside lying parallel to each other was heated until the enamel insulation evaporated (the glass was slightly incandescent) and then pinched twice very quickly with fine nose pliers (see Fig. 3, II and III). The probe tubes were sterilized in the same manner as the plain glass tubes in the autoclave.

The fine copper and constantan wires used to make the thermocouple gave a rapid response to temperature change because the speed of response of a thermocouple is inversely proportional to its mass. The conduction of heat to the couple along the length of the wires was minimized by their small size. To check the amount of heat conduction and the speed of response, preliminary observations using a bare couple

in a water bath were made. These showed that when the couple was moved even  $1/8$  to  $1/16$  inch out of the water, with the leads still in the water, that its temperature dropped immediately. Hence, the fact that the leads to the couple were in contact with the side of the tube had no bearing on the reading given by the probe. In other words, the probe accurately reflected the temperature and changes in that temperature which pertained to the medium in which the couple was situated. For the above reasons, it is believed that the thermocouple accurately measured the temperature of the milk at any given instant bearing in mind the 2-second full scale speed of the recorder used.

The length of time involved in the heating and cooling of the samples of milk in the glass tubes was estimated from the moving chart during the heating process. Because heating occurred in one water bath, and cooling took place in an ice water bath beside it, the length of time that the milk was exposed in the heating bath was only controllable to the nearest two seconds. After the experiment, the exact length of time could be measured from the chart with the use of dividers and a ruler.

To aid in the measurement of the 16 seconds that was aimed for, an "L" shaped piece of wire was



fastened to the recording pen so that it extended a distance equivalent to 16 seconds in the direction that the chart paper was moving. Thus, when the place where the recording line had crossed the minimum temperature line on the chart was reached by the wire, the milk had been held at or above that temperature for sixteen seconds and was a signal to lift the tubes out of the hot water bath and plunge them into the ice water for cooling.

#### Enumeration

Counts on the material in the tubes after pasteurization were made according to Standard Methods ( 2 ) with the following modifications:

In order to remove the milk from the sealed glass tubes, it was necessary to break them open. This was done in the region of the air cushion left between the milk and the end of the tube sealed after the milk was placed in it. First, the glass was nicked or scratched using a three sided file. Then the nick and the outside of the glass were wiped with tissue soaked in methyl alcohol, the end of the tube and the nick quickly flamed, and the glass broken at the nick with a flamed pair of needle nose pliers. A twist of the wrist gave a clean break each time. The milk was then tapped into a sterilized dilution

blank giving a 1:100 dilution of the contents of the tube. It was sometimes necessary to swing the blank and the tube in an arc to completely drain the tube. To facilitate the removal of the milk in any future modification, bending the tubes into a flattened "U" shape with an air cushion in each arm would allow the tube to be opened at both ends allowing the milk to drain more easily out of the narrow bore.

TABLES AND FIGURES

Table 1

## MEAN STANDARD PLATE COUNTS OF COMMERCIAL MILK

Plant	Tank	Raw Plate Count	Pasteurized Plate Counts	
			Immediate	After Storage*
1.	1	$4.5 \times 10^7$	$2.9 \times 10^5$	$1.9 \times 10^5$
	2	$5.3 \times 10^7$	$4.0 \times 10^5$	$2.6 \times 10^5$
	3	$7.6 \times 10^6$	$1.3 \times 10^5$	$7.0 \times 10^4$
2.	1	$1.2 \times 10^5$	$3.7 \times 10^2$	$4.2 \times 10^1$
	2	$1.5 \times 10^5$	$6.5 \times 10^2$	$2.1 \times 10^2$
	3	$3.1 \times 10^4$	$4.9 \times 10^2$	$1.7 \times 10^2$
1.	1	$1.9 \times 10^7$	$1.0 \times 10^5$	$6.5 \times 10^4$
	2	$3.6 \times 10^7$	$2.1 \times 10^5$	$1.5 \times 10^5$
	3	$6.1 \times 10^6$	$1.6 \times 10^5$	$1.2 \times 10^5$
3.	1	$2.0 \times 10^7$	$5.9 \times 10^4$	$8.7 \times 10^4$
	2	$1.2 \times 10^7$	$5.6 \times 10^4$	$9.2 \times 10^4$
	3	$1.5 \times 10^6$	$5.2 \times 10^4$	$5.2 \times 10^4$
4.**	1	$7.4 \times 10^5$	$7.4 \times 10^4$	$5.1 \times 10^4$
	2	$8.4 \times 10^5$	$5.1 \times 10^4$	$4.3 \times 10^4$
4.**	1	$2.2 \times 10^6$	$9.1 \times 10^4$	$5.1 \times 10^4$
	2	$3.9 \times 10^6$	$9.9 \times 10^4$	$5.4 \times 10^4$
4.**	1	$2.9 \times 10^6$	$8.7 \times 10^4$	$7.0 \times 10^4$
	2	$2.5 \times 10^6$	$1.3 \times 10^5$	$1.0 \times 10^5$

\* 10 days at 4 - 5°C. (39.2 - 41°F.)

\*\* Milk from Plant No. 3 processed in 320 lb. lots at  
Dept. of Food Science, University of Manitoba

Table 2

## MEAN PSYCHROPHILIC COUNTS OF COMMERCIAL MILK

<u>Plant</u>	<u>Tank</u>	<u>Raw Plate Count</u>	<u>Pasteurized Plate Counts</u>	
			<u>Immediate</u>	<u>After Storage*</u>
1.	1	$7.6 \times 10^6$	$1.3 \times 10^1$	$3.3 \times 10^0$
	2	$3.8 \times 10^7$	$9.1 \times 10^0$	$8.3 \times 10^0$
	3	$2.1 \times 10^6$	$7.5 \times 10^0$	$4.1 \times 10^0$
2.	1	$3.0 \times 10^5$	$7.5 \times 10^0$	0.0
	2	$2.3 \times 10^5$	$1.6 \times 10^1$	0.0
	3	$5.0 \times 10^3$	0.0	$1.6 \times 10^0$
1.	1	$8.8 \times 10^5$	$4.1 \times 10^0$	$2.5 \times 10^0$
	2	$1.0 \times 10^6$	0.0	$3.1 \times 10^0$
	3	$1.5 \times 10^5$	$1.0 \times 10^1$	$1.6 \times 10^0$
3.	1	$2.3 \times 10^7$	$1.2 \times 10^0$	$1.4 \times 10^4$
	2	$1.5 \times 10^7$	$6.6 \times 10^{-1}$	$6.4 \times 10^3$
	3	$1.9 \times 10^6$	$1.4 \times 10^0$	$3.7 \times 10^3$
4.**	1	$9.0 \times 10^5$	$1.1 \times 10^0$	0.0
	2	$1.3 \times 10^6$	$5.0 \times 10^{-1}$	0.0
4.**	1	$2.9 \times 10^6$	$1.2 \times 10^0$	0.0
	2	$1.1 \times 10^6$	$1.4 \times 10^0$	0.0
4.**	1	$1.6 \times 10^6$	$8.8 \times 10^{-1}$	0.0
	2	$2.9 \times 10^6$	$1.8 \times 10^0$	0.0

\* 10 days at 4 - 5°C. (39.2 - 41°F.)

\*\* Milk from Plant No. 3 processed in 320 lb. lots at Dept. of Food Science, University of Manitoba

Table 3

PLATE COUNTS OF PSYCHROPHILIC STRAINS OF  
Pseudomonas fluorescens

<u>Strain</u>	<u>Trial</u>	<u>Before Pasteurization</u>		<u>After Pasteurization</u>	
		<u>Counting Chamber Estimate</u>	<u>Plate Count</u>	<u>Immediate Plate Count</u>	<u>After Storage** Plate Count</u>
3756	1	$1.98 \times 10^9$	-	Negative	200
		$1.98 \times 10^{11}$	-	"	100
		$1.98 \times 10^{10}$	-	"	100
	2	$1.98 \times 10^7$	-	"	200
		$1.98 \times 10^5$	-(L.A.)	"	$1.87 \times 10^5$ *
9428	1	$2.33 \times 10^8$	$1.5 \times 10^7$	"	0
		$2.33 \times 10^6$	$2.9 \times 10^5$	"	0
		$2.33 \times 10^4$	-	"	0
	2	$2.33 \times 10^9$	$3.3 \times 10^8$	"	0
		$2.33 \times 10^5$	$2.9 \times 10^4$	"	0
10038	1	$2.60 \times 10^9$	$3.7 \times 10^8$	"	500
		$2.60 \times 10^7$	$9.1 \times 10^6$	"	900*
		$2.60 \times 10^7$	$9.1 \times 10^6$	"	4200
	2	$2.60 \times 10^8$	$8.4 \times 10^7$	"	950
		$2.60 \times 10^4$	$9.6 \times 10^3$	"	3800*
		$2.60 \times 10^4$	$9.6 \times 10^3$	"	2500
3756	1	$3.29 \times 10^8$	$6.0 \times 10^8$	"	0
		$3.29 \times 10^2$	$3.0 \times 10^2$	"	0
		$3.29 \times 10^5$	$6.3 \times 10^5$	"	0

\*\* 10 days at 4 - 5°C. (39.2 - 41.0°F.)

\* Tube with thermocouple leads

(L.A.) = leak past thermocouple leads

Table 3 - Continued

<u>Strain</u>	<u>Trial</u>	<u>Before Pasteurization</u>		<u>After Pasteurization</u>	
		<u>Counting Chamber Estimate</u>	<u>Plate Count</u>	<u>Immediate Plate Count</u>	<u>After Storage Plate Count</u> **
3756	2	$3.29 \times 10^6$	$6.3 \times 10^6$	Negative	0
		$3.29 \times 10^3$	$6.0 \times 10^3$	"	0
		$3.29 \times 10^3$	$6.0 \times 10^3$	"	0
9428	1	$1.27 \times 10^9$	$7.8 \times 10^8$	"	150
		$1.27 \times 10^9$	$7.8 \times 10^8$	"	0 *
		$1.27 \times 10^6$	$6.3 \times 10^5$	"	0
	2	$1.27 \times 10^7$	$1.0 \times 10^7$	"	0
		$1.27 \times 10^3$	$8.0 \times 10^2$	"	0
		$1.27 \times 10^3$	$8.0 \times 10^2$	"	0 *

\*\* 10 days at 4 - 5°C. (39.2 - 41.0°F.)

\* Tube with thermocouple leads

# Fig. I HTST Pasteurization

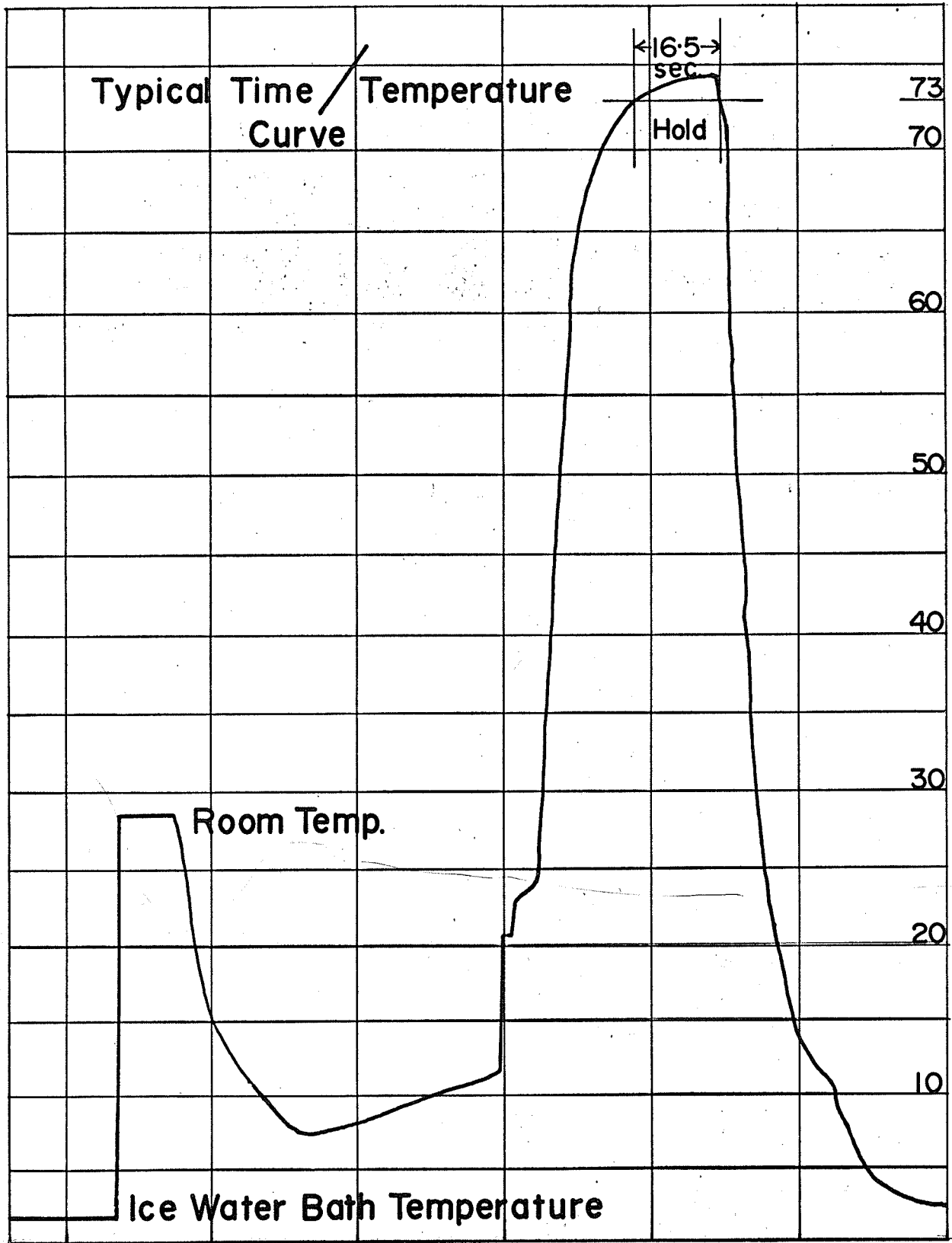
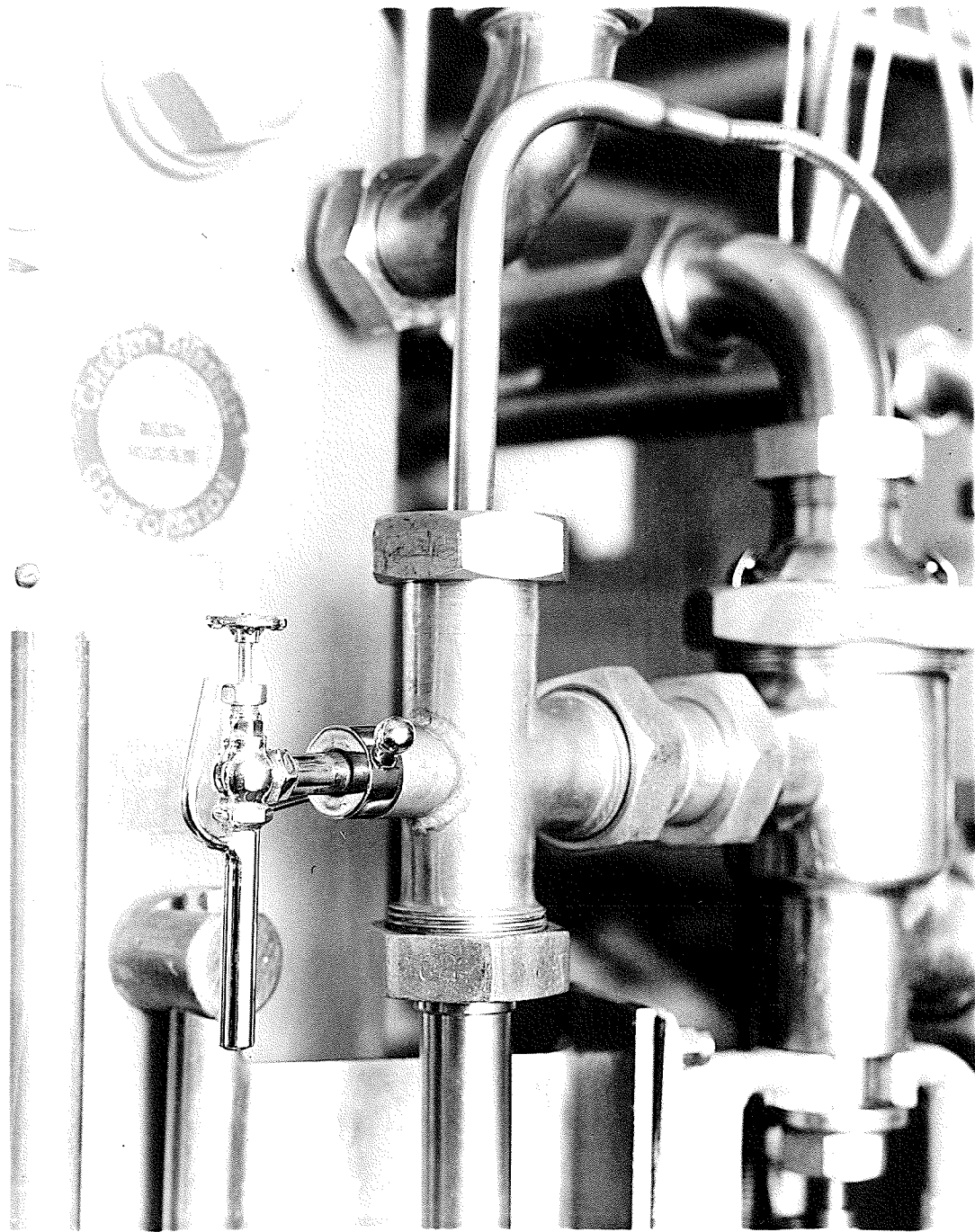


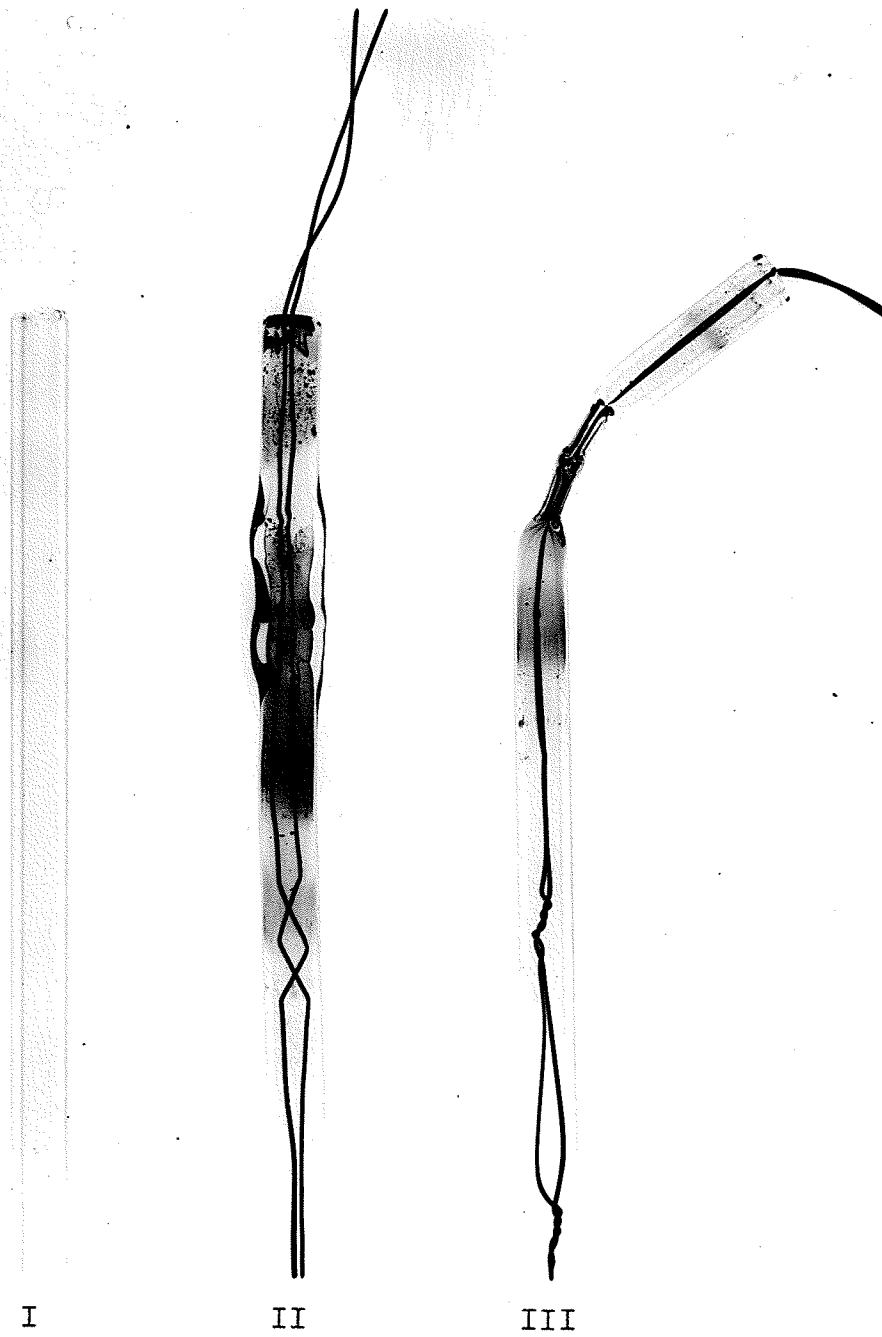


Figure 2



APV SIDE-ARM SAMPLING VALVE - on HTST pasteurizer

Figure 3



GLASS TUBES FOR LABORATORY PASTEURIZATION x 1.5  
I - flint tube, 5 mm. O.D., 3 mm. I.D.  
II & III - thermocouple leads in temperature  
sensing tube

RESULTS AND DISCUSSION

## Results and Discussion

The psychrophilic bacteria that limit the keeping quality of pasteurized milk have generally been assumed to be the result of post-pasteurization contamination. Macaulay et al (56) gave four psychrophilic strains of *Pseudomonas fluorescens* an HTST pasteurizing treatment in 5 ml. of milk and found survivors in highly significant numbers after storage for 10 days at 3 - 5°C. If psychrophilic strains are thermoduric with regard to present HTST pasteurizing treatments, then serious reassessment of raw milk's bacterial quality is needed. To show whether Macaulay's work was of any practical commercial significance, the first experimental series was proposed.

The original hypothesis was that pasteurized milk taken aseptically from the end of the holding tube of an HTST pasteurizer contains microorganisms capable of growth in refrigerated milk. Because the numbers of these organisms would be very small initially, a period of growth at refrigerated temperatures in the milk was recognized as necessary.

For this hypothesis to be valid, demonstrable changes would have to occur during the refrigerated period. The following standards were proposed for acceptance: 1. Standard Plate Count (SPC) should

show an increase between samples counted immediately and those held for 10 days at 4 - 5°C. 2. Psychrophilic Plate Counts should be able to show both the presence and numbers of the bacteria that grow at 4 - 5°C.

The first experiment used four different HTST units and duplicate samples were removed from the ends of their holding tubes. One duplicate was given SPC and psychrophilic plate count soon after pasteurization and the other held at 4 - 5°C. for 10 days before SPC and psychrophilic plate counts were made again. Macaulay also held his sample 10 days but at 3 - 5°C. Because there was no way of choosing between bacteria from the milk's natural flora which had merely lain dormant at 4 - 5°C. for 10 days and those that had actively grown in it during this time, incubation of the psychrophilic plates was also at 4 - 5°C. This is in contrast to Macaulay who could incubate his plates at 20°C. since he dealt with a single psychrophilic strain. Results of this present study took longer to be evident.

In the second experiment, the glass tubes were used first, to eliminate any chance of any particle of milk, as well as its pasteurizing container, not receiving the full HTST heat treatment; and second,

to eliminate all chance of post-pasteurization contamination during the 10 day storage period. In contrast, Macaulay used stainless steel tubes closed with rubber stoppers for heating and cooling containers. For the 10 days storage, the milk had to be transferred to glass test-tubes.

In Table 1, the results from 18 tank lots of commercial milk are summarized and compared using the Standard Plate Count. The raw, post-pasteurized, and after storage pasteurized counts show: the effect of pasteurization in the change between raw and pasteurized; and the effect of the 10 days storage in the change between pasteurized and the after storage pasteurized counts. In Table 2, these same 18 tank lots are similarly summarized and compared using a psychrophilic plate count.

The key counts are those made after 10 days storage. These samples were not opened after being taken from the end of the holding tube. Rogick and Burgwald (81), (80) examined samples immediately after HTST pasteurization and found no psychrophiles. On re-examination after 7 days at 4 - 7°C., these same samples all showed evidence of growth. This was attributed to psychrophiles. By using a duplicate un-opened tube, possible contamination was stopped.

Tables 1 and 2 show no general pattern of growth during the 10 day storage in either SPC or psychrophilic counts. They also show rather surprisingly high counts in both raw and pasteurized milk. These may be due to the warmer months in which this work was done. Large numbers of psychrophilic bacteria are to be noted in the raw milk from Table 2 in company with similar high counts by SPC for the same milk in Table 1.

Plant No. 3 shows increases. The chief cause was a refrigeration failure in the cold storage room where plates and samples were stored. A blown fuse stopped the compressors and allowed samples and plates to reach 15°C. (60°F.) before discovery and correction. These results show the importance of continuous low storage temperatures to prevent growth of bacteria able to grow at temperatures just above those that allow psychrophiles to grow. Thus even brief removal of milk containers from cold storage to a warm environment has its effect and allows growth.

In none of the three tank lots processed at Plant No. 3 did the psychrophilic count reach levels likely to cause objectionable flavor as indicated by Boyd et al (9).

When milk from Plant No. 3 was later brought

to the University Food Science Department for processing at 72.2°C. (162°F.), the legal minimum, there were no significant increases. Instead, the mean counts declined.

Discussing similar work, Olsen (65) points out that processing plants with good records have less psychrophile trouble. Plant No. 2 is an example. Here, well organized sanitizing, operating, and clean up applied by trained workers is checked by in-plant and external laboratories. Their low raw counts result from continuous conscientious effort by field staff on the farm coupled with bonuses for low raw milk counts.

How much influence has the raw SPC on subsequent keeping quality? It has been suggested that high count raw milk is more suited to psychrophilic growth after pasteurization. Overcast and Adams (67) investigated this theory and found no stimulation for Ps. fluorescens. They did find stimulation for a Brevibacterium lipolyticum and one other psychrophile isolate. An inhibitory effect on Ps. fragi was shown after 1, 2, and 4 days. Their conclusion, the same as Erdman and Thornton's (26), (27), was that psychrophiles in milk are a sanitation problem.



The results for experiment 2 are shown in Table 3. The immediate post pasteurization counts are all zero. The counts for the majority made 10 days after pasteurization are below the limit of 30 per plate using a 1:100 dilution. The first trial used Ps. fluorescens strain 3756 and the four tubes had mean counts of 2, 1, 1, and 2 respectively. When a trial using this organism was repeated, the after storage counts were uniformly zero.

The results using Ps. fluorescens strain 10038 show survivors after storage. This shows that some organisms were able to survive the heat treatment. Macaulay (56) consistently found survivors immediately after pasteurization with all three organisms. Macaulay's pre-pasteurization counts ranged from approximately 200,000 to more than 1,000,000. This is the chief difference between the results of the two laboratory investigations. This explains why Macaulay's counts after 10 days storage were so much higher than those in this study. The recording chart for the first trial of 10038 shows an exposure of 16 sec. at 73°C. or above, with maximum temperature reached 73.5°C., and a bath temperature of 74°C. During the second trial of 10038, the maximum temperature was 1°C. lower. No effect is evident from this.

Comparison of HTST time - temperature treatment in Exp. 2 with a commercial HTST shows that the glass tube method took about 26 seconds to reach 73°C. (162°F.). Franklin (32) indicates 40 seconds for this in a HTST unit. Cooling was abrupt from 73°C. as shown in Figure 1. Thus this method gave considerably less heat treatment than normal HTST units at temperatures below that of the 73°C. - 16 second treatment.

The sealed glass tube method's chief advantage was its absolute guarantee of freedom from contamination during the 10 day incubation at 4 - 5°C. A further advantage was the exact record of heat treatment from the thermocouple tube. The semi-micro nature of the tubes gave rapid temperature change with simple and inexpensive apparatus.

Disadvantages encountered with the method were: awkward removal of milk from the tubes to dilution blanks, the anaerobic conditions in the sealed tube, and the problem of giving identical exposures to separate lots. The first could be remedied by having longer Pasteur pipettes. Mechanical devices might overcome the third since this depends, in part, on the eye-hand co-ordination of the operator in the transfer from heating to cooling bath. That the

sealed tube is much different from conditions at the bottom of a full milk bottle or carton is doubtful. Sinclair and Stokes (86) found higher yields of organisms from liquid media at lower temperatures due to the greater amount of oxygen dissolved. Most modern HTST units have vacuum sections for flavor removal that also remove air and oxygen. Because of autoclaving and mixing by inverting rather than shaking, little air was likely to be incorporated in the milk prior to pasteurizing in the glass tubes. Thus, there is little to indicate any difference between the oxygen tension in the sealed glass tubes, the screw cap tubes used in Exp. 1, and the ordinary milk container.

Upadhyay and Stokes (97) state that as incubation temperature is lowered, lag phase of growth as well as exponential, stationary, and death phases are lengthened. If this is the reason for no growth in either experiment during the 10 day period at 4 - 5°C., then the keeping quality problem is one of maintaining uniformly low temperatures. This suggests that taking samples in the same way as Experiments 1 and 2, incubating at 15 - 20°C. for 2 - 3 days would speed up the lag phase. Incubating psychrophilic plates at 4 - 5°C. for 15 - 20 days would follow. This was not done in this study because Macaulay's

results indicated survivors without it. If, when starting with natural populations, survivors from such a procedure are found, then, either these are facultative psychrophiles, or the higher incubation temperature has allowed recovery of cold temperature growth ability.

Possibly the heat shock of the HTST treatment in Experiment 2 caused damage that only a few isolated cells were able to overcome. Perhaps the damage was such that the medium used in plating did not meet the more demanding nutritional needs of the heat shocked bacteria. Of course, it could also be that the milk itself no longer met these increased nutritional needs and the bacteria died. If so, then they didn't survive pasteurization.

A possible survival hypothesis is that a few bacteria repair their heat damaged enzyme systems and other disorganization in the cell. As with all chemical and cellular processes, low temperatures would slow this repair. Higher temperatures within their growth range might allow a recovery of low temperature growth ability through such repair being done faster.

From the results of these experiments, Macaulay's contention that psychrophiles are

thermoduric is entirely possible. However, its significance relative to practical HTST pasteurization is open to question because the occurrence of large enough numbers of any one particular thermoduric organism in raw milk is unlikely. Thus, isolated cases of psychrophiles surviving commercial HTST pasteurization are possible, but the more likely cause of psychrophiles in pasteurized milk is post-pasteurization contamination.

CONCLUSIONS

### Conclusions

In Experiment 1, natural populations of psychrophilic bacteria were found in commercial raw milk using psychrophilic plate counts. In milk taken from the ends of HTST holding tubes, these same bacteria failed to be detected in any significant amount, even after 10 days incubation in milk at 4 - 5°C. as indicated by ability to grow on solid media at this same temperature.

In Experiment 2, pasteurization of selected strains of psychrophilic Ps. fluorescens in high concentrations by an HTST treatment in 3 mm. bore glass tubes failed to show any pattern of survival generally but with a specific organism a low survival rate was observed on a solid medium at 4 - 5°C.

From these two experiments, it is concluded that psychrophilic growth in milk after pasteurization is not likely to be caused by survivors of pasteurization but by post-pasteurization contamination.

REFERENCES



## REFERENCES

1. American Public Health Association - Standard Methods for the Examination of Dairy Products Microbiological and Chemical, 10th Edition, New York, 1953.
2. American Public Health Association - Standard Methods for the Examination of Dairy Products, 11th Edition, New York, 1960.
3. Andrey, J. Jr., and Frazier, W.C. - PSYCHROPHILES IN MILK HELD TWO DAYS IN FARM BULK COOLING TANKS - J. Dairy Sci. 43:1774 - 1782. 1960.
4. Anonymous - NEWS AND EVENTS - J. Milk and Food Tech. 23:59. 1960.
5. Atherton, H.F., Doan, F.J. and Watrous, G.H. Jr., - CHANGES IN BACTERIAL POPULATION AND CHARACTERISTICS OF BOTTLED MARKET MILK DURING REFRIGERATED HOLDING - Bull. Pa. Agric. Expt. Sta. No. 575, 1954. Abstract - Dairy Sci. Abstr. 16:654C. 1954.
6. Baumann, D.P., and Reinhold, G.W. - THE ENUMERATION OF PSYCHROPHILIC MICROORGANISMS IN DAIRY PRODUCTS - J. Milk and Food Tech. 26:351 - 356. 1963.
7. Bonner, M.D. and Harnom, L.G. - CHARACTERISTICS OF ORGANISMS CONTRIBUTING TO SPOILAGE IN COTTAGE CHEESE \* J. Dairy Sci. 40:1599 - 1611. 1957.
8. Boyd, J.C., Smith, C.K., and Trout, G.M. - THE EFFECT OF THE INCUBATION TIME AND TEMPERATURE ON THE DETERMINATION OF THE PSYCHROPHILIC BACTERIA IN MILK BY THE AGAR PLATE METHOD - J. Milk and Food Tech. 17: 365 - 368. 1954
9. Boyd, J.C., Smith, C.K., and Trout, G.M. - THE ROLE OF PSYCHROPHILIC BACTERIA IN THE KEEPING QUALITY OF COMMERCIALY PASTEURIZED AND HOMOGENIZED MILK - J. Milk and Food Tech. 18: 32 - 36. 1955
10. Broitman, S., Mallmann, W.L., and Trout, G.M. - A SIMPLE TEST FOR DETECTING KEEPING QUALITY IN MILK - J. Milk and Food Tech. 21:280 - 284. 1958.

11. Burgwald, L.H., and Josephson, D.V. - THE EFFECT OF REFRIGERATOR STORAGE ON THE KEEPING QUALITIES OF PASTEURIZED MILK - J. Dairy Sci. 30:371 - 383. 1947.
12. Castell, C.H., and McDermott, L.A. - MULTIPLICATION OF BACTERIA IN WATER AND ITS SIGNIFICANCE IN FOOD SPOILAGE - Food Res. 7: (3) 244 - 253. 1942.
13. Chaudhary, R.A., Tuckey, S.L., and Witter, L.D. - HEAT RESISTANCE OF THREE STRAINS OF PSYCHROPHILIC ORGANISMS ADDED TO SKIM-MILK FOR COTTAGE CHEESE MANUFACTURE - J. Dairy Sci. 43: 1774 - 1782. 1960.
14. Collins, E.B. - FACTORS INVOLVED IN THE CONTROL OF GELATINOUS CURD DEFECTS OF COTTAGE CHEESE. I STORAGE TEMPERATURE AND PH - J. Milk and Food Tech. 18: 169 - 171. 1955.
15. Collins, E.B. - FACTORS INVOLVED IN THE CONTROL OF GELATINOUS CURD DEFECTS OF COTTAGE CHEESE. II INFLUENCE OF PH AND TEMPERATURE UPON THE BACTERICIDAL EFFICIENCY OF CHLORINE - J. Milk and Food Tech. 18: 189 - 191. 1955.
16. Corley, R.T. and Hammer, B.W. - BACTERIOLOGICAL STUDIES ON CREAMERY WATER SUPPLIES - Abstract of Paper presented at 37th Ann. Meeting, - J. Dairy Sci. 25: 723 - 724. 1942.
17. Craige, Capt. J.E. - SIGNIFICANCE OF CONCENTRATION OF COLIFORM ORGANISMS IN RAW MILK UPON SURVIVAL OF PASTEURIZATION - J. Milk and Food Tech. 9:191 - 196. 1946.
18. Csenge, J.L. and Doan, F.J. - THE NEOTETRAZOLIUM TEST FOR DETERIORATION OF STORED CREAM, CONCENTRATED MILK AND COTTAGE CHEESE - J. Milk and Food Tech. 21: 223 - 225. 1958.
19. Davis, P.A., and Babel, F.J., - SLIME FORMATION ON COTTAGE CHEESE - J. Dairy Scie.37: 176 - 184. 1954.
20. Day, E.A. and Doan, F.J. - A TEST FOR THE KEEPING QUALITY OF PASTEURIZED MILK - J. Milk and Food Tech. 19: 63 - 66. 1956.

21. Druce, R.G., and Thomas, S.B. - THE MICROBIOLOGICAL EXAMINATION OF BUTTER - J. Applied Bacteriol. 22: 52 - 56. 1959.
22. Elliker, P.R. - A WAY TO BETTER KEEPING QUALITY OF DAIRY PRODUCTS - Milk Dealer 52: (10):40 - 41, 74 - 76. 1963.
23. Elliker, P.R., Sing, E.L., Christenson, L.J., and Sandine, W.E. - PSYCHROPHILIC BACTERIA AND KEEPING QUALITY OF PASTEURIZED DAIRY PRODUCTS - J. Milk and Food Tech. 27: 69 - 75. 1964.
24. Enright, J.B., Sadler, W.W., and Thomas, R.C. - PASTEURIZATION OF MILK CONTAINING THE ORGANISM OF "Q" FEVER - Amer. Jour. Publ. Health 47: (b) 695 - 700. 1957.
25. Enright, J.B. - THE PASTEURIZATION OF CREAM, CHOCOLATE MILK AND ICE CREAM MIXES CONTAINING THE ORGANISM OF "Q" FEVER - J. Milk and Food Tech. 24: 351 - 355. 1961.
26. Erdman, I.E., and Thornton, H.E. - PSYCHROPHILIC BACTERIA IN EDMONTON MILK AND CREAM, I. NUMBERS - Can J. Technol. 29: 232 - 237. 1951.
27. Erdman, I.E., and Thornton, H.R. - PSYCHROPHILIC BACTERIA IN EDMONTON MILK AND CREAM, II. KINDS - Can. J. Technol. 29: 238 - 242. 1951.
28. Evans, D.A., Lachman, E.L., and Litsky, W. - SOME OBSERVATIONS ON THE BACTERIOLOGICAL KEEPING QUALITY OF MILK PROCESSED BY HIGH TEMPERATURES WITH A 0.6 SECOND HOLDING TIME - J. Milk and Food Tech. 26: 332 - 336. 1963.
29. Fabian, F.W. - THERMODURIC ORGANISMS IN RELATION TO HIGH TEMPERATURE, SHORT-TIME PASTEURIZATION - J. Milk Tech. 5: 237 - 242. 1942.
30. Ford, H.F., and Babel, F.J. - MILK QUALITY PROBLEMS ASSOCIATED WITH PRESENT DAY MARKETING - J. Milk and Food Tech. 22: 141 - 144. 1959.

31. Foster, H.G. Jr., Lear, S.A., and Metzger, H.J. - TIME-TEMPERATURE STUDIES OF THE INACTIVATION RATE OF Brucella abortus STRAIN 2308 IN MILK - J. Milk and Food Tech. 16: 116 - 120. 1953.
32. Franklin, J.G. - A SIMPLE LABORATORY-SCALE HTST MILK PASTEURIZER - J. Dairy Res. 32: 281 - 289. 1965.
33. Freeman, T.R., Nanavati, N.V., and Glenn, W.E. - FASTER ENUMERATION OF PSYCHROPHILIC BACTERIA IN PASTEURIZED MILK - A PRELIMINARY REPORT - J. Milk and Food Tech. 27:304 - 307. 1964.
34. Galesloot, T.E. - (English title) THE INFLUENCE OF HEAT TREATMENT ON THE KEEPING QUALITY OF PASTEURIZED MILK (De invloed van de pasteurisatie-temperatuur op de duurzaamheid van gepasteuriseerde melk) - Neth. Milk Dairy Jour. 9 (4):237 - 248. 1955. Abstract - Dairy Sci. Abstr. 18:427. 1956.
35. Gilcreas, F.W., and O'Brien, J.E. - THERMAL DEATH RANGE OF BACTERIA IN MILK (A NEW ELECTRIC SAMPLING DEVICE) - J. Milk and Food Tech. 9: 269 - 272. 1946.
36. Glazier, L.R. - QUALITY COMPARISONS OF HIGH-TEMPERATURE, SHORT-TIME AND ULTRA-HIGH-TEMPERATURE PASTEURIZED MILK - J. Milk and Food Tech. 26: 347 - 351. 1963.
37. Greene, V.W., and Jezeski, J.J., - INFLUENCE OF TEMPERATURE ON THE DEVELOPMENT OF SEVERAL PSYCHROPHILIC BACTERIA OF DAIRY ORIGIN - Applied Microbiol. 2: 110 - 117. 1954.
38. Harmon, L.G. - COTTAGE CHEESE PROBLEMS IN PRODUCTION AND SANITATION - QUALITY CONTROL IN COTTAGE CHEESE - J. Milk and Food Technol. 26: 86 - 90. 1963.
39. Heather, C.D., and Vanderzant, W.C., - EFFECTS OF TEMPERATURE AND TIME OF INCUBATING AND PH OF PLATING MEDIUM ON ENUMERATING HEAT-TREATED PSYCHROPHILIC BACTERIA - J. Dairy Sci. 40:1079 - 1086. 1957.
40. Heather, C.D., and Vanderzant, W.C. - EFFECT OF THE PLATING MEDIUM ON THE SURVIVAL OF HEAT-TREATED CELLS OF Pseudomonas fluorescens - Food Res. 22:164 - 169. 1957.

41. Hempler, P. - PSYCHROPHILIC BACTERIA IN PASTEURIZED MILK - Nord.Vetmed. 7: (9) 705 - 746. 1955. Abstract - Dairy Sci. Abstr. 18:244. 1956.
42. Hileman, J.L., and Leber, H. - HIGH TEMPERATURE, SHORT-TIME PASTEURIZATION AND ITS PRACTICAL APPLICATION TO THE DAIRY INDUSTRY - J. Milk and Food Tech. 4: 128 - 137. 1941.
43. Hileman, J.L., Leber, H., and Speck, M.L. - THERMODURIC BACTERIA IN PASTEURIZED MILK II. STUDIES ON THE BACTERIA SURIVING PASTEURIZATION, WITH SPECIAL REFERENCE TO HIGH-TEMPERATURE, SHORT-TIME PASTEURIZATION - J. Dairy Sci. 24: 305 - 315. 1941.
44. Holland, R.F. and Dahlberg, A.C. - A STUDY OF THE TIME-TEMPERATURE RELATIONSHIPS IN THE PASTEURIZATION OF MILK AS REGARDS CREAMING, PHOSPHATASE AND BACTERIAL DESTRUCTION - J. Dairy Sci. 23: 504 - 505. 1940.
45. Holmquist, C.A. and Tiedeman, W.D. - ENGINEERING OF PASTEURIZATION - J. Milk Tech. 1 (1):11 - 15.(Oct.) 1937.
46. Ingraham, J.L., and Stokes, J.L. - PSYCHROPHILIC BACTERIA - Bacteriol. Rev. 23: 97 - 108. 1959.
47. Jezeski, J.J., and Macy, J. - CRYOPHILIC ORGANISMS IN WATER AND BUTTER - J. Dairy Sci. 29: 439 - 452. 1946.
48. Kastli, P., and Binz, M. - (English title)KEEPING QUALITY OF PASTEURIZED MILK IN COLD STORES (Die Halborkeit von pasteurisierter Flaschenmilch bei Kuhlagerung) - Schweiz Milkchztg. 82 ( 5 Wissenschaftl. Beilage Nr. 34) 265 - 272 (1956)(Fed. Inst. Dairy Ind. Bact., Liebefeld, Berne, Switzerland). Abstract - Dairy Sci. Abstr. 18:428. 1956.
49. Kaufmann, O.W. and Andrews, R.H. - THE DESTRUCTION RATE OF PSYCHROPHILIC BACTERIA IN SKIMMILK - J. Dairy Sci. 37: 317 - 327. 1954.
50. Kennedy, L. and Weiser, H. - SOME OBSERVATIONS ON BACTERIA ISOLATED FROM MILK THAT GROW WITHIN A PSYCHROPHILIC TEMPERATURE RANGE - J. Milk and Food Tech. 13:353 - 357. 1950

51. Kolmer and Boirner - Approved Laboratory Technic, 4th Edition, 1945 D. Appleton-Century Co. - as quoted in - DIRECTIONS FOR USE OF HELBER AND PETROFF-HAUSER COUNTING CHAMBERS FOR STANDARDIZATION OF VACCINES - published by C.A. Hausser and Son, Philadelphia, Pa.
52. Kreuger, P.F. - THE INTRODUCTION INTO CHICAGO OF THE HIGH-TEMPERATURE SHORT TIME METHOD OF MILK PASTEURIZATION - J. Milk Tech. 1 (7):29 - 33. (Nov.) 1938.
53. Kreuger, P.F. - SHORT TIME PASTEURIZATION. B. A SANITARIAN'S VIEW - Ass. Bull. (Int.Ass. Milk Dealers) No. 13, pp 334 - 340. 1939. Abstract - Dairy Sci. Abstr. 1:216. 1939.
54. Lawton, W.C. and Nelson, F.E. - THE EFFECT OF STORAGE TEMPERATURES ON THE GROWTH OF PSYCHROPHILIC ORGANISMS IN STERILE AND LABORATORY PASTEURIZED SKIMMILKS - J. Dairy Sci. 37: 1164 - 1172. 1954.
55. Leesment, H. and Dufeu, J. - A RAPID METHOD FOR DETERMINING THE PSYCHROPHILIC COUNT IN MILK - Int. Dairy Congr. 1962 C, pp 392 - 398. 1962.
56. Macaulay, D.M. - THE EFFECT OF PASTEURIZATION ON THE SURVIVAL OF CERTAIN PSYCHROPHILIC BACTERIA - Thesis M - 1187, University of Manitoba, 1962.
57. Marth, D.H., and Frazier, W.C. - BACTERIOLOGY OF MILK HELD AT FARM BULK COOLING TANK TEMPERATURES III. PSYCHROPHILES FOUND AND THEIR GROWTH - J. Milk and Food Tech. 20: 93 - 99. 1957.
58. Martin, W.H., Foltz, V.D., and Rutz, W.D. - A SURVEY OF COTTAGE CHEESE QUALITY - J. Milk and Food Tech. 23: 306 - 310. 1960.
59. Mickelson, R., Fish, Nancy L., and Claydon, T.J. - CHEMICAL AND FLAVOR CHARACTERISTICS OF A MILK PROTEOLYSATE OF Pseudomonas fluorescens - J. Dairy Sci. 48: 783. 1965.
60. Morris, C.S. - CRYOPHILIC BACTERIA AS A CAUSE OF MILK SAMPLES FAILING THE METHYLENE BLUE TEST - Dairy Industr. 7: (3) 63 - 64, 69. 1942.

61. Nelson, F.E. and Baker, M.P. - INFLUENCE OF TIME AND TEMPERATURE OF PLATE INCUBATION UPON BACTERIAL COUNTS OF MARKET MILK AND RELATED PRODUCTS, PARTICULARLY AFTER HOLDING UNDER REFRIGERATION - J. Milk and Food Tech. 17: 95 - 100. 1954.
62. Oginsky, E.L. and Umbreit, W.W. - An Introduction to Bacterial Physiology - W.H. Freeman and Co. Inc., San Francisco. 1959.
63. Olsen, R.H. and Jezeski, J.J. - SOME EFFECTS OF CARBON SOURCE, AERATION AND TEMPERATURE ON GROWTH OF A PSYCHROPHILIC STRAIN OF Pseudomonas fluorescens - J. Bacteriol. 86: 429 - 433. 1963.
64. Olson, J.C., Jr., Macy, H., and Halvorson, H.O. - THERMAL DEATH TIME STUDIES OF COLIFORM BACTERIA IN MILK - J. Dairy Sci. 32: 702. 1949.
65. Olson, J.C., Jr., Willoughby, D.S., Thomas, E.L., and Morris, H.A. - THE KEEPING QUALITY OF PASTEURIZED MILK AS INFLUENCED BY THE GROWTH OF PSYCHROPHILIC BACTERIA AND THE ADDITION OF AUREOMYCIN - J. Milk and Food Tech. 16: 213 - 219. 1953
66. Olson, J.C., Jr., Parker, R.B., and Mueller, W.S. - THE NATURE SIGNIFICANCE AND CONTROL OF PSYCHROPHILIC BACTERIA IN DAIRY PRODUCTS - J. Milk and Food Tech. 18: 200 - 203. 1955.
67. Overcast, W.W. and Adams, G.A. - GROWTH OF CERTAIN PSYCHROPHILIC BACTERIA IN PASTEURIZED MILK AS INFLUENCED BY PREVIOUS EXCESSIVE PSYCHROPHILIC GROWTH IN THE RAW MILK - J. Milk and Food Tech. 29: 14 - 18. (Jan.) 1966.
68. Overcast, W.W. and Skean, J.D. - GROWTH OF CERTAIN LIPOLYTIC MICROORGANISMS AT 4°C. AND THEIR INFLUENCE ON FREE FAT ACIDITY AND FLAVOR OF PASTEURIZED MILK - J. Dairy Sci. 42: 1479 - 1485. 1959.
69. Parfitt, E.H. - SOME BACTERIOLOGICAL PROBLEMS INVOLVED IN HTST PASTEURIZATION - Rep. N.Y. St. Ass. Dairy and Milk Insp., pp. 195 - 219. 1939. Abstract - Dairy Sci. Abstr. 2: 143. 1939.
70. Parker, R.B., Smith, V.N., and Elliker, P.R. - BACTERIA ASSOCIATED WITH A GELATINOUS OR SLIMY CURD DEFECT OF COTTAGE CHEESE - J. of Dairy Sci. 34: 887 - 893. 1951.

71. Parker, R.B., Coldwell, A.L., and Elliker, P.R. - PSYCHROPHILIC BACTERIA - A SANITATION PROBLEM - J. Milk and Food Tech. 16: 136 - 139, 152. 1953.
72. Post, F.J., and Krishnamurty, G.B. - SUGGESTED MODIFICATIONS OF THE CALCIUM ALGINATE SWAB TECHNIQUE - J. Milk and Food Tech. 27: 62 - 65. 1964.
73. Prouty, C.C. - THE EFFECTS OF HEAT ON BACTERIA IN MILK WITH PARTICULAR REFERENCE TO THERMAL DEATH RATES AS INFLUENCED BY VARIOUS FACTORS - J. Milk and Food Tech. 16: 66 - 72. 1953.
74. Prouty, C.C. - OBSERVATION ON THE PRESENCE OF FACULTATIVE PSYCHROPHILIC BACTERIA IN MILK PRODUCED UNDER THE FARM TANK SYSTEM - J. Milk and Food Tech. 18: 250 - 253. 1955.
75. Prucha, M.J. and Parfitt, E.H. - HIGH TEMPERATURE PASTEURIZATION - Manual Dairy Manuf. Conf., Univ. Illinois, pp. 213 - 220. 1938. Abstract - Dairy Sci. Abstr. 1: 339. 1938.
76. Punch, J.D., Olson, J.C., Jr., and Thomas, E.L. - PRELIMINARY OBSERVATION ON POPULATION LEVELS OF PURE CULTURES OF PSYCHROPHILIC BACTERIA NECESSARY TO INDUCE FLAVOR OR PHYSICAL CHANGE IN PASTEURIZED MILK - J. Dairy Sci. 44: 1160. 1961. (Abstract)
77. Punch, J.D. and Olson, J.C., Jr. - COMPARISON BETWEEN STANDARD METHODS PROCEDURE AND A SURFACE PLATE METHOD FOR ESTIMATING PSYCHROPHILIC BACTERIA IN MILK - J. Milk and Food Tech. 27: 43 - 47. 1964.
78. Randolph, H.E., Freeman, T.R., and Petersen, R.W. - KEEPING QUALITY OF MARKET MILK OBTAINED AT RETAIL OUTLETS AND AT PROCESSING PLANTS - J. Milk and Food Tech. 28: 92 - 96. 1965.
79. Robinson, H.B. - PRESENT PROBLEMS IN HIGH HEAT PASTEURIZATION PROCESSES - J. Milk and Food Tech. 23: 49 - 53. 1960.
80. Rogick, F.A., and Burgwald, L.H. - A STUDY ON PSYCHROPHILIC BACTERIA IN MARKET MILK - J. Dairy Sci. 33: 403. 1950.



81. Rogick, F.A. and Burgwald, L.H. - SOME FACTORS WHICH CONTRIBUTE TO THE PSYCHROPHILIC BACTERIAL COUNT IN MARKET MILK - J. Milk and Food Tech. 15: 181 - 185. 1952.
82. Roth, N.G., and Wheaton, R.B. - CONTINUITY OF PSYCHROPHILIC AND MESOPHILIC GROWTH CHARACTERISTICS IN THE GENUS Arthrobacter - J. Bacteriol. 83: 551 - 555. 1962.
83. Schutt, D.B. - THE ROLE OF NON-FAECAL BACTERIA IN WATER SUPPLIED TO CREAMERIES - Can. Jour. Pub. Health 27: 226 - 227. 1936.
84. Sherman, J.M., Cameron, G.M., and White, J.C. - THE BACTERIOLOGICAL SPOILAGE OF MILK HELD NEAR THE FREEZING POINT - Abst. of Paper presented to Ann. Meeting Amer. Dy. Sci. Assoc., June 1941. J Dairy Sci. 24: 526 - 527. 1941.
85. Sinclair, N.A., and Stokes, J.L. - FACTORS WHICH CONTROL MAXIMAL GROWTH OF BACTERIA - J. Bacteriol. 83: 1147 - 1154. 1962.
86. Sinclair, N.A. and Stokes, J.L. - ROLE OF OXYGEN IN THE HIGH CELL YIELD OF PSYCHROPHILES AND MESOPHILES AT LOW TEMPERATURES - J. Bacteriol. 85: 164 - 167. 1963.
87. Speck, M.L. - THE INACTIVATION OF BACTERIA IN MILK EXPOSED TO ULTRA-HIGH PASTEURIZATION TEMPERATURES - J. Milk and Food Tech. 24: 358 - 361. 1961.
88. Straka, R.P. and Stokes, J.L. - METABOLIC INJURY TO BACTERIA AT LOW TEMPERATURES - J. Bacteriol. 78: 181 - 185. 1959.
89. Straka, R.P. and Stokes, J.L. - PSYCHROPHILIC BACTERIA FROM ANTARCTICA - J. Bacteriol. 80: 622 - 625. 1960.
90. Stumbo, C.R. - BACTERIOLOGICAL CONSIDERATIONS RELATING TO PROCESS EVALUATION - Food Tech. 2: 115 - 132. 1948.
91. Sulzbacher, W.L. - SURVIVAL OF MICROORGANISMS IN FROZEN MEAT - Food Technol. 4: 386 - 390. 1950.
92. Thomas, S.B. - PSYCHROPHILIC MICRO-ORGANISMS IN MILK AND DAIRY PRODUCTS - Dairy Sci. Abstracts 20: 355 - 370, 447 - 468. 1958.

93. Thomas, S.B., Hobson, P.M., and Bird, E.R. - PSYCHROPHILIC BACTERIA IN MILK - Int. Dairy Cong. 1959, 3: 1334 - 1340. 1959.
94. Thomas, S.B., Griffiths, J.M., and Foulkes, J.B. - THE PSYCHROPHILIC COLONY COUNT OF MILK - Dairy Eng. 78: 49 - 51, 61. 1961.
95. Thomas, S.B., Thomas, B.F., and Ellison, D., - MILK BACTERIA WHICH GROW AT REFRIGERATOR TEMPERATURES - Dairy Inds. 14: 921 - 925, 946. 1949.
96. Thome, K.E., and Ljunggren, B. - PSYCHROPHILIC BACTERIA IN MARKET MILK - (psykrofila bakterier i konsumtionmjolk) - Rep. Milk and Dairy Res. Alnarp, Sweden 52 (1957). Abstract - Dairy Sci. Abstracts 20 (6): 511. 1958.
97. Upadhyay, J. and Stokes, J.L. - ANAEROBIC GROWTH OF PSYCHROPHILIC BACTERIA - J. Bacteriol. 83: 270 - 275. 1962.
98. Upadhyay, J. and Stokes, J.L. - TEMPERATURE-SENSITIVE FORMIC HYDROGENLYASE IN A PSYCHROPHILIC BACTERIUM - J. Bacteriol. 85: 177 - 185. 1963.
99. Upadhyay, J. and Stokes, J.L. - TEMPERATURE-SENSITIVE HYDROGENASE AND HYDROGENASE SYNTHESIS IN A PSYCHROPHILIC BACTERIUM - J. Bacteriol. 86: 992 - 998. 1963.
100. VanDerZant, W.C. and Moore, A.V. - THE INFLUENCE OF CERTAIN FACTORS ON THE BACTERIAL COUNTS AND PROTEOLYTIC ACTIVITIES OF SEVERAL PSYCHROPHILIC ORGANISMS - J. Dairy Sci. 38: 743 - 750. 1955.
101. Wagenaar, R.O. and Jezeski, J.J. - THE INFLUENCE OF THE TYPE OF DILUENT ON THE GROWTH AND SURVIVAL OF Pseudomonas putrefaciens - J. Dairy Sci. 35: 738 - 754. 1952.
102. Weber, C.W. - SAFETY FACTORS OF HIGH TEMPERATURE SHORT TIME PASTEURIZERS - J. Milk and Food Tech. 10: 14 - 26, 53. 1947.
103. Workman, T.W. - SHORT TIME, HIGH TEMPERATURE PASTEURIZATION - Ass. Bulletin (Int. Ass. Milk Dealers) 33: 585. 1941. Abstract - Dairy Sci. Abstracts 3: 264. 1941-42.