

Dep. Col.  
Thesis  
M 139

DEPOSITORY  
COLLECTION  
NOT TO BE  
TAKEN

THE UNIVERSITY OF MANITOBA  
LIBRARY

ACCESSION NUMBER

51046

A THESIS

entitled

"A Biochemical Investigation of the Distribution of  
Glucosides in Plants Indigenous to Northwest Canada  
and adjacent territories".

presented to

The University of Manitoba

by

D. Roy McCullagh

as partial requirement for the Master of Science Degree.

April 24, 1926.

CONTENTS

Part One.

(1) Introduction.

- (a) Present status of our knowledge of glucosides.
- (b) The principle of the "Biochemical Method".
- (c) Aim of the present work.

(2) Detailed account of method and technique.

- (a) Preparation of the plant extract.
- (b) Biochemical hydrolysis.
- (c) Clarification.
- (d) Estimation of the optical activity.
- (e) Estimation of the reducing power.
- (f) The enzymes, and their preparation.

(3) The enzymolytic reduction index.

Part Two.

(1) General account of work done.

(2) Detailed account of work done with discussion of results.

- A. Assays in which the saccharimeter was used.
- B. Assays in which the polarimeter was used.

Part Three.

(1) Summary of results.

(2) Bibliography.

PART ONE

(1) Introduction.

(a) Present status of our knowledge of glucosides.

In the year 1901 Emile Bourquelot introduced enzymes into the laboratory as reagents(1). This marked an epoch in the study of glucosides and their distribution in plants, as previous to that time no systematic work had been done on these substances although ten glucosides had been discovered in a more or less accidental manner, e.g. salicin(2). The method of studying glucosides and their distribution devised by this noted savant has since become known as "The Biochemical Method of Bourquelot". The efficacy of the method is shown by the fact that out of 281 species of phanerogames examined in his laboratory, previous to the year 1920, 205 were found to contain one or more beta-glucosides(3). The method has been used by many French workers to demonstrate the presence of known glucosides in a large number of new plants, and to indicate the presence of previously unknown glucosides in a number of plants. This led to the isolation of a number of new glucosides.

(b) The principle of the "Biochemical Method".

The "Biochemical Method of Bourquelot" is based on the principle of enzymolytic hydrolysis. The distribution of sucrose is studied by the use of invertase; that of beta-glucosides by means of emulsin; and that of glucosides not

effected by the enzymes mentioned above by the use of "fermenting powders" prepared from the plants under consideration. The effect of these enzymes on plant extracts is measured by observing the changes, during hydrolysis, in the optical activity and reducing power. The result is qualitative, in that the ratio between the change in the reducing power and the change in the optical activity is specific for each glucoside; and the result is also quantitative, in that the total amount of the reducing sugar formed by the complete hydrolysis in an extract of known concentration gives the measurement of the glucosides present.

(c) Aim of the present work.

In the present work this method has been applied to a number of plants indigenous to the northwest part of this continent. The choice was only made after a phylogenetic consideration of those species found to be positive by the French workers. Thus it is purposed:-

1. To discover whether or not plants on this continent have the same glucosidic content as related species in Europe, and
11. To demonstrate the possible presence of new glucosides as a preliminary step towards their isolation and detailed investigation. These are all of potential pharmacological value in that they usually represent some of the most active compounds of the plant, i.e. hormones etc. The knowledge of

these compounds is of far reaching importance in the study of plant physiology. "The recognition of the potent effect of the constituents of glucosides in acting as stimuli and starters of active metabolism may be of importance in studying the nutrition of animals".(4)

(2) Detailed account of methods and technique.

(a) Preparation of the plant extract.

The fresh plant tissue is placed in a proportionate quantity of boiling 85% alcohol in a reflux apparatus, boiled for thirty minutes, filtered, minced, and again extracted in the same manner. The combined extracts are evaporated under reduced pressure, and taken up with sufficient thymol water to make one cubic centimeter of the extract represent one gram of the fresh material.

(b) Biochemical hydrolysis.

Invertase is added to two of three flasks each containing 25cc. of the extract. After incubation for one week at 37 C. emulsin is added to one of the flasks containing the invertase and incubation is continued for another week. The result is, one control flask, one flask in which the sucrose has been hydrolysed, and one flask in which the sucrose and beta-glucosides (if present) have been hydrolysed. In that the emulsin of almonds usually contains invertase, it is necessary to hydrolyse with invertase first even in those cases where only the beta-glucosides are to be studied.

(c) Clarification.

General method: A saturated solution of neutral lead acetate is added to each flask until no more precipitate is formed, and brought to equal volume with water. The precipitate is filtered off, the excess lead precipitated with hydrogen sulphide and removed by filtration. Aliquot quantities of the filtrate are neutralised with a concentrated solution of ammonium hydroxide, made up to equal volume with water, treated with animal charcoal by boiling under a reflux and filtered.

Frequently this is insufficient and special treatment must be devised to bring about satisfactory clarification.

(d) Estimation of the optical activity.

Three drops of ammonium hydroxide are added to each flask to stabilize any sugars and glucosides present. As shown by H. Colin and A. Chaudin (5) this is very essential in that the addition of  $\text{NH}_4\text{OH}$  during the reaction rapidly increases the rotatory power of beta-glucosides and reduces that of alpha-glucosides to the limit of rotation. After these changes are complete the optical activity of each is measured and expressed as degrees (with  $l=2$  dm.)

The first part of the work herein reported was done with a saccharimeter (F. Schmidt and Haensch no. 7765), which was only graduated to 0.1% glucose. In spite of careful standardization this portion of the work suffered much on account of the inaccuracy of this instrument. The latter

part of the work was done with a "Duboseq-Pellin Precision Model" polarimeter of great accuracy.

(e) Estimation of reducing power.

The method of Benedict(6) was used exclusively in preference to that of Bertrand (7), used by most of the French workers. The latter method is more tedious, requires a larger amount of solution than is available when the reducing power is low, and gives identical results. The former method, however, requires an absolute constancy of technique in order to attain the same accuracy.

The reducing power is expressed as percentage glucose.

(f) The enzymes and their preparation.

As will be readily understood it is essential that the invertase used in the above procedure be entirely free from beta-glucosidase. If this is not the case the addition of emulsin will bring about no further change and the method is valueless for the estimation of beta-glucosides. The invertase in the animal intestine is present in such small quantities that the preparation of a large amount was found to be very tedious. Yeast is the best source of invertase, so for this purpose a strain must be found that contains no beta-glucosidase. Top fermenting yeast from Shea's Brewery Ltd. reached this requirement, and was the only satisfactory strain of three tried. The method used by Bourquelot (8) to prepare



the invertase was found to be unsatisfactory, as all the enzymes and co-enzymes necessary for alcoholic fermentation are present, with the result that much of the sugar is destroyed before the necessary estimations can be made. The method of C.S. Hudson (9) consists of cytolysing by means of toluene and distilled water, precipitation of some impurities with lead acetate, removal of lead with hydrogen sulphide, and removal of acetic acid by dialysis. This is very satisfactory except that a solid preparation is more convenient in that it obviates change in volume. Such a solid preparation is very readily obtained by precipitating Hudson's solution with 2.5 volumes of 95% alcohol; decanting off the supernatant liquid after 16 hours; filtering or centrifuging; washing thoroughly with alcohol and ether to remove all traces of water; and drying in vacuo. If not free from water the precipitate, on coming in contact with the air, becomes dark brown instead of remaining perfectly white. This preparation has the advantage over all commercial preparations seen by the author in that it gives no color to an aqueous solution, and thus precludes the necessity of clarifying a solution which is otherwise clear.

The emulsin was prepared, by the method of Bridel and Arnold (10), from almonds and was entirely satisfactory.

The preparation of invertase is extremely active, 50 mg. being sufficient to change the optical activity of 25cc.

of an 8% sucrose solution to zero in ninety minutes. There is no perceptible loss in activity within eight months. The emulsin is also quite active; 62 mg. will change ten cc. of a 5% solution of amygdalin to a rotation of zero in two hours.

(3) The enzymolytic reduction index.

The ratio  $\frac{\text{increase in reducing power (measured as mg. glucose)}}{\text{change in optical activity (degrees with } l=2 \text{ dm.)}}$  is known as the enzymolytic reduction index, and is specific for each pure glucoside. This has been calculated for sucrose and a large number of beta-glucosides by the French chemists. In using this index when working on extracts of plant tissues it must be remembered that if one obtains the index for a known glucoside it does not necessarily signify the presence of that glucoside, and vice versa, as it may be complicated by a number of factors. For example there may be two or more glucosides present and the combined indices give the result obtained. Also in the estimation of the optical activity of the original solution the glucoside itself may not be present, having been removed by the process of clarification, and unless the index for the pure glucoside has been obtained after the same process of clarification the indices will not be comparable. On the other hand if two closely related plants give the same index it can be assumed with a moderate degree of safety that their glucosidic content is the same.

PART TWO(1) General account of work done.

During the summer of 1925 fifteen species of plants were collected in sufficient quantity for preliminary survey, and the initial treatment carried to a stage at which the extracts could be preserved without decomposition.

Estimations have been completed on certain of the plant tissues for twelve of these species: the remainder, including certain of the various tissues separately extracted in some of those species already studied, are now being examined.

(2) Detailed account of work done with discussion of results.A. Assays in which the saccharimeter was used.1. Salix interior--Rowlee. (willow)

This specimen was identified by Mr. J. F. Higham of the Manitoba Agricultural College; it was collected on June 17th and extracted on the following day.

It has been known for many years that the bark of different species of *Salix* contains a beta-glucoside, viz. salicin(2). The bark of *Salix interior* is shown to contain this substance by the following results.

	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Index.</u>
After invertase.	+0.31°	0.710%	
After emulsin.	+0.65°	0.830%	353

In that the index for salicin is 321, these results are plainly indicative of its presence.

2. Aralia nudicaulis L. (wild sarsaparilla)

This plant was collected on June 9th and identified by Mr. J. F. Higham. The following results were obtained from an extract of the roots made the day of collection.

	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Index</u>
Original.	+0.27°	1.176%	
After Invertase.	-0.01°	1.363%	667
After emulsin.	±0.00°	1.357%	---

The enzymolytic reduction index for sucrose is 603, which so approximates that found in the above determinations that the presence of that sugar is almost certain. There is no significant change after incubation with emulsin, which is the result anticipated, in that so many of the closely related species contain glucosides not hydrolysed by this enzyme. *Hedera helix* is known to contain the flavone glucoside rutin. Sarsaparilla of many varieties are known to contain saponins, e.g. Honduras sarsaparilla(11).

3. Asarum canadense, L. (wild ginger)

This collection was made at Victoria Beach on June 13th, identified by Mr. G. W. Lowe of the University of Manitoba, and extracted the same day. Estimations were made on the various parts of the plant with the following results:

<u>Leaves.</u>	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Indices</u>
Original.	+0.68°	0.287%	
After invertase.	+0.27°	0.625%	824
After emulsin.	+0.28°	0.740%	---

<u>Stems.</u>	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Indices.</u>
Original.	+0.40°	0.114%	
After invertase.	+0.26°	0.175%	435
After emulsin.	+0.21°	0.186%	---
<u>Roots.</u>			
Original.	+0.42° (?)	0.184%	
After invertase.	+0.27°	0.422%	---
After emulsin.	+0.27°	0.500%	---

In that there is a large experimental error, on account of the instrument, in measuring the optical activity the results of the hydrolysis by invertase are taken by the author to be a demonstration of the presence of sucrose in all the tissues examined. The changes in the reducing power after emulsin indicate the presence of a glucoside in every case. The accompanying changes in optical activity could not be demonstrated with the instrument in use, and were therefore probably rather small, which means that the index for the glucoside must be quite large.

4. Diervilla diervilla, (L.) MacM. (bush honeysuckle)

This plant was identified by Mr. C. W. Lowe on June 13th, collection and extraction was carried out on the same day.

<u>Root; 1st assay</u>	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Indices</u>
Original	+1.06°	0.280%	
After invertase.	+0.08°	0.741%	470
After emulsin.	+0.73°	1.250%	783

<u>2nd assay.</u>	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Indices</u>
Original.	+0.60°	0.523%	
After invertase.	+0.02°	0.833%	534
After emulsin.	+0.71°	1.316%	700
<u>Stems:1st assay.</u>			
Original	+0.41°	0.549%	
After invertase.	-0.06°	0.840%	831
After emulsin.	+0.21°	1.087%	914
<u>2nd assay.</u>			
Original	+0.32°	0.388%	
After invertase.	±0.00°	0.654%	619
After emulsin.	+0.22°	1.(?)%	

In every assay there is definite evidence of the presence of sucrose. The indices vary considerably from that of sucrose (603) but are all within the limit of experimental error. Hydrolysis with emulsin definitely proves the presence of a beta-glucoside in the roots and stems. The index does not correspond to that of any of the known glucosides and suggests the probable presence of a glucoside hitherto not isolated.

B. Assays for which the polarimeter was used.

1. Cypripedium parviflorum, Salisb. (Downey Lady's Slipper)

This plant was gathered near Birds' Hill, Manitoba, on June 19th, and extracted on the same day. It was identified by Miss Grace Cameron of the University of Manitoba.

The flowers, leaves and stems were separately examined. In the case of the flowers only 19 grams of the fresh material was available to make the necessary 50cc. of solution for the study of beta-glucosides alone. This lcc. of the extract represents only 0.38 gms. of the fresh material instead of the orthodox 1.0 gms.

The results of the estimations on the flowers are:

	<u>Optical Activity</u>	<u>Reducing Power</u>
After invertase.	+0.06°	0.877%
After emulsin.	+0.62°	0.703%

These results indicate that the emulsin brought about a synthesis, which used some of the reducing sugar previously present. It is rather confounding in that the optical activity changed to the right. One would expect either the formation of some beta-glucoside or gentiobiose, but if this had been the case the rotation would have been changed to the left.

The following results were obtained from the leaves:

	<u>Optical Activity</u>	<u>Reducing Power.</u>	<u>Indices.</u>
Original.	+2.37°	0.574%	
After invertase.	-0.27°	2.415%	697
After emulsin.	+0.15°	2.415%	25

The enzymolytic reduction index obtained from the changes after the use of invertase indicates that the greater part of the substance hydrolysed by that enzyme is probably sucrose (index 603). The fact that it is higher than the index for the pure sugar probably indicates the presence of some such substance such as stachyose or raffinose (cf. results on stems of same species).

The changes in reducing power after emulsin could not be detected. As the experimental error in this determination is sometimes as high as 0.5% these results indicate the presence of a beta-glucoside with an enzymolytic reduction index in the vicinity of 25. The only index corresponding to this is that of verbenalin, which is 19.(12).

<u>Stems.</u>	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Indices</u>
Original.	+0.55°	1.659%	
After invertase.	-0.15°	2.155%	737
After emulsin.	-0.44°	2.475%	-1103

These results are of special interest in that the optical activity is changed to the left after the use of emulsin. For this reason the enzymolytic reduction index is designated as negative.

It will be remembered that raffinose is hydrolysed by invertase to form fructose and melibiose; also stachyose to form fructose and mannotriose. In both cases the change in rotation to the left is less than during the hydrolysis of sucrose, both because there is less fructose formed and because the remaining substances are more highly dextro-rotatory than glucose. This would result in an increase for the enzymolytic reduction index for invertase, as is the case here. The results of the estimations after emulsin indicate the possibility of one or other of these substances being present, as the hydrolysis of either melibiose or mannotriose by emulsin changes the optical activity to the left. The enzymolytic reduction index is about that which would be expected from meli-



There is the possibility that these results indicate the presence of some glucoside formed from a laevorotatory sugar instead of glucose.

It was expected that this plant might contain the glucoside loroglossin (index 407) as do many species of the Order Orchidaceae(13). Obviously this is not the case.

2. Pulsatilla patens(L.), Mill. (Prairie crocus)

<u>Flowers.</u>	<u>Optical Activity</u>	<u>Reducing Power</u>
After invertase.	- 2.00°	5.980%
After emulsin.	- 2.36°	5.828%

These results are comparable to those obtained in the case of the flowers of *Cypripedium parviflorum*, in that there is an enzymic synthesis. These however are more readily explained in that the formation of a beta-glucoside or of gentiobiose would change the rotation to the left as has occurred.

Foliage and Stems

This estimation included all the plant above the ground except the colored portion of the flower.

	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Indices.</u>
Original.	+ 2.34°	2.140%	
After invertase.	+0.81°	3.185%	683
After emulsin.	+0.63°	3.448%	-1463

These results are almost identical with those of the stems of *Cypripedium parviflorum* and the same conclusions are to be drawn, i.e. the presence of certain polysaccharides or of a glucoside from a laevorotatory sugar.

<u>Roots.</u>	<u>Optical Activity.</u>	<u>Reducing Power.</u>	<u>Indices.</u>
Original.	+0.93°	0.142%	
After invertase.	-0.04°	0.851%	731
After emulsin.	+0.21°	1.234%	1532

These results point towards the presence of sucrose and one or more beta-glucosides. The index of enzymolytic reduction after emulsin is not similar to those of any of the beta-glucosides known and indicates the presence of a glucoside, which is probably still to be isolated.

### 3. Rhus Toxicodendron L. (Poison Ivy)

This specimen was gathered by Doctor M. S. Hollenberg at Grand Beach on June 20th. The extract was made on June 23rd, and was strongly toxic.

<u>Leaves.</u>	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Index</u>
After invertase.	-1.03°	0.934%	
After emulsin.	-1.21°	0.956%	-122

Again we find a change to the left in optical activity accompanied by an increase in the reducing power. The significance of this has been mentioned in connection with similar results in this report.

### 4. Prunus pennsylvanica L. (pin cherry)

The ripe fruit had been gathered three days previous to extraction; the following results were obtained from an extract of the epicarp and mesocarp, i.e. the skin and flesh of the fruit.

	<u>Optical Activity</u>	<u>Reducing Power</u>
Original.	-2.06°	6.6(?)%
After invertase.	-2.02°	6.754%
After emulsin.	-1.94°	6.873%

These changes are too small to make the indices of any value but indicate the presence of sucrose and a beta-glucoside.

5. *Padus virginiana*, (L) Mill. (choke cherry)

The extract in this case also was from the epicarp and mesocarp, and made two days after collection.

	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Indices</u>
Original.	- 2.93°	11.200%	
After invertase.	- 2.92°	11.111%	---
After emulsin.	-2.68°	11.235%	517

These results indicate the absence of a compound hydrolysed by invertase, and the presence of more than a trace of one or more beta-glucosides. The index of enzymolytic reduction is exactly that of prunasin so that in all probability that is the glucoside present. Prunasin has been isolated from the bark of *Prunus serotina*(14), which is possibly the same species and from the young branches of *Cerasus Padus* (15). Prulaurasin (index 359) has been isolated from *Prunus lauroceracus*(16), and amygdalin is widely distributed amongst all the Rosacea. It is possible that the index in this determination is an indication of the presence of this latter glucoside which has an index of 490.

6. Commandra pallida, A.D.C.

The plant was identified by Mr. C. W. Lowe, collected at Victoria Beach on June 13th, and extracted the same day. Experimental results obtained from the stem are:

	<u>Optical Activity</u>	<u>Reducing Power</u>
Original.	-0.26°	0.094%
After invertase.	-0.26°	0.177%
After emulsin.	-0.32°	0.195%

The results on this hemiparasite are of interest because of their negativity. There are only traces of substances present that are hydrolysed by either invertase or emulsin. In that the plant has chlorophyl one would expect that a certain amount of anabolism should take place in the tissues. That such is the case is shown by the presence of these traces of substances hydrolysed by these enzymes. That the plant probably derives a great deal of its nourishment from its host seems probable from the fact that these substances are there in such small quantity.

7. Pyrola sp. (wintergreen)

This was identified by Miss Grace Cameron, collected on June 19th near Birds Hill, and extracted the same day. On examination of the stems and leaves the following results were obtained:

	<u>Optical Activity</u>	<u>Reducing Power</u>	<u>Indices</u>
Original	-1.72°	1.234%	
After invertase.	-0.46°	1.372%	-133
After emulsin.	+0.10°	1.470%	175

These results are the only ones in which the rotation is changed to the right after the use of invertase. For this reason the index is designated as negative. This is a very peculiar but interesting result. Hydrolysis by means of emulsin demonstrates the presence of a beta-glucoside, with an index which does not correspond to that of any of the known glucosides. The closely related species assayed by other workers contain such glucosides as monotropitin(17), monotropein(18) and gaultherin(19), which are not hydrolysed by emulsin.

8. Symphocarpus racemosus, Michx. (snowberry)

The plant was identified by Mr. C. W. Lowe, at Victoria Beach; the berries were collected on September 13th and extracted on the 14th with the following results:

	<u>Optical Activity.</u>	<u>Reducing Power</u>	<u>Indices.</u>
Original.	-0.24°	5.152%	
After invertase.	-1.49°	5.835%	546
After emulsin.	-0.88°	6.170%	549

Sucrose is present as shown by the changes after invertase. The index 549(after emulsin) is midway between that of syringin (570) and that of prunasin (517).

9. Symphocarpus occidentalis, Hook. (wolfberry)

Mr. J. F. Higham identified the plant which was gathered and extracted on September 10th. In this species the clarification was not sufficiently complete to allow polarimetric

estimation. An extract of the berries showed an increase from 1.818% to 9.216% after the use of invertase, and a further increase to 9.568% after hydrolysis with emulsin. This indicates the probable presence of a large amount of sucrose and one or more beta-glucosides.

### PART THREE

#### Summary of results.

One or more glucosides hydrolysable by emulsin were found in some tissue of eleven of the thirteen species examined. The only species giving entirely negative results was *Aralia nudicaulis* in which only the roots have been examined, and which is related to many species containing glucosides not hydrolysed by emulsin. In three species the index indicates the probable presence of glucosides previously identified in similar species of other workers, and in the remaining species the glucosides are possibly not yet known.

In certain of the assays the results indicate either the presence of certain of the polysaccharides, or glucosides of laevorotatory sugars not previously known to exist naturally in glucosidic combination.

Sucrose was demonstrated in nearly all the tissues examined with the use of invertase, except in the case of some of the fruits, where the sugar content seemed to be chiefly in the form of monosaccharides.

Enzymic synthesis was demonstrated in the case of two of the flowers examined.

The author wishes to express his appreciation for the kindly assistance extended to him by Professor A. T. Cameron and Dr. F. D. White, of the department of Biochemistry, under whose supervision the work was carried out; by Professor A. H. Reginald Buller in selection of species; and by various members of the departments of Botany at the University and the Agricultural College in collection and identification of the plants.

The author is also indebted for the use of a polariscope purchased by Professor A. T. Cameron with a grant from the National Research Council of Canada.

BIBLIOGRAPHY.

1. Bourquelot: J.Pharm.Chim.,1901,14,481.
2. Leroux: Ann,de Chim.et de Phys.,2<sup>e</sup> serie,t41,p 295,1829.
3. Em.Bourquelot: Comptes Rendu,171,p 473.
4. E.F.Armstrong: The Carbohydrates and Glucosides,p.230.
5. H. Colin and A.Chaudun: Bull,Soc,Chim.,1924,35,p 974.
6. Benedict: Jour.Am. Med. Ass'n.,1911,57, 1193.
7. Bertrand: Bull.Soc.Chim.de France,1906,35,128.
8. Abderhalden: Handbuch der Biochemischen Arbeitsmethoden,Bd.7,p<sup>767</sup><sub>76</sub>
9. C. S. Hudson: J.Am.Chem.Soc.,1914,p1566, ~~XXXVI~~.
10. Bridel et Arnold: Bull.Soc.Chim.Biol.,Tome 11,p 216.
11. H.P.Kaufman und C.Fuchs: Ber.,1923,56B,p 2527.
12. L. Bourdier: Archiv der Pharmazie, 1908,Bd,246,S.275.
- 13a.Bourquelot et Bridel: Journ.Pharm.et Chim.,1914,X,p14.
  - b.Ibid: 1919, XX,p 81.
  - c.M.P.Delauney: Bull.Soc.Chim.Biol.,Tome iii,p 208.
  - d.Ibid: Tome V,p 398.
14. F.B.Power and C.W.Moore: J.Chem.Soc.,1909,95,p 243.
15. Herissey: Arch.Pharm.,1907,245,p 641.
16. Ibid: 1907,245,p 473.
17. Marc Bridel: Bull.Soc.Chim.Biol.,Tome V,p 918.
18. Ibid: Tome V, p 722.
19. E.F.Armstrong: The Carbohydrates and Glucosides, p 195.