- 1 Predators and alate immigration influence the season-long dynamics of
- 2 soybean aphid (Hemiptera: Aphididae)
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Abstract

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The identification of key species within multi-predator assemblages suppressing agricultural pests is paramount to the incorporation of predators into pest management plans. Generalist predators have strong impacts on population growth of soybean aphids (Aphis glycines, Hemiptera: Aphidae) in North America, but their relative influence may vary seasonally, and potentially interact with immigration of alate aphids following initial field colonization. Here we present the results of a season-long study in Minnesota that used cage manipulations to estimate weekly impacts of predation on field populations of soybean aphids, while concurrently quantifying alate aphid and predator densities. We used generalized linear models based on field level aphid and predator counts, and aphid counts on caged and uncaged experimental plants, to show that *Harmonia axyridis* (Coleoptera: Coccinellidae) was negatively associated with A. glycines abundance, even under alate immigration levels associated with aphid outbreaks, suggesting potential regulation of aphid populations. In contrast, two other common predators, Coccinella septempunctata (Coleoptera: Coccinellidae) and Orius insidiosus (Hemiptera: Anthocoridae), did not show patterns of association with aphid dynamics. We also show that A. glycines populations only have significant positive associations with the number of immigrating alatae on uncaged experimental plants that had higher alate densities than the field average. The negative effect of H. axyridis on aphid populations was also observed on uncaged experimental plants, suggesting that even high levels of alate aphid immigration did not disrupt predator suppression. These results add to the growing body of evidence demonstrating that even low numbers of certain generalist predators can effectively suppress pest populations in agroecosystems. In particular, our findings suggest that the abundance of both H. axyridis and alate aphids are critical to soybean aphid seasonal dynamics.

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Keywords: alatae; *Aphis glycines*, generalist predators; *Harmonia axyridis*; predator-prey dynamics

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- **Highlights:**
- Harmonia axyridis abundance was negatively associated with Aphis glycines abundance
- Aphids did not vary with abundance of Coccinella septempunctata or Orius insidiosus
- High levels of alate immigration did not disrupt aphid suppression by predators

• Generalist predators provided effective season-long aphid suppression in soybean

1. Introduction

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causing mortality to pests in agricultural ecosystems and this pest control yields significant economic benefits (Landis et al., 2008; Losey and Vaughan, 2006; Schipanski et al., 2016; Zhao et al., 2016). There is considerable interest in determining the impacts of specific natural enemy guilds on pest suppression, particularly for newly introduced pests (Zhao et al., 2016). Though much of the focus in the past was on classical biological control, there has more recently been interest in quantifying the impact of naturally occurring natural enemies on pests, and in developing strategies to augment or otherwise improve these valuable ecosystem services (i.e. conservation biocontrol) (Landis et al., 2000; Symondson et al., 2002). This shift in focus has renewed interest in examining the characteristics of specific natural enemies that make them effective in suppressing pest populations. Traditionally, 'ideal' biocontrol agents were considered to be species with high prey specificity, since these 'specialists' have mechanisms to efficiently seek out particular prey/hosts and will not be diverted by the presence of alternative prey (Murdoch et al., 1985; Symondson et al., 2002). These assertions originate, at least partially, from early theoretical work that assumed the ideal outcome of biocontrol efforts was low, stable equilibrium populations of the biocontrol agent and pest (Murdoch, 1994; Murdoch et al., 1985). There is a growing body of both theoretical and experimental literature that demonstrates that generalist predators can act as effective biocontrol agents in agricultural ecosystems (Chang and Kareiva, 1999; Murdoch et al., 1985; Stiling and Cornelissen, 2005; Welch et al., 2012), and that low stable equilibrium populations are not a strict requirement for effective, long-term pest suppression (Murdoch et al., 1985; Symondson et al., 2002). Generalist insect predators represent an important component of the natural enemy assemblage associated with insect pests (Bannerman et al., 2015; Symondson et al., 2002). Here we use the term generalist predators to include both polyphagous species with a wide prey range and stenophagous taxa that are opportunistic feeders on various resources (e.g. Coccinellidae) (Hodek and Evans, 2012; Welch et al., 2012). Common predators of insect pests in agricultural habitats includes beetles (Coleoptera: Coccinellidae & Carabidae), lacewings (Neuroptera: Chrysopidae), true bugs (Hemiptera: Anthocoridae, Nabidae, & Reduviidae), flies (Diptera:

Syrphidae) and arachnids (Araneae) (Pedigo and Rice, 2009; van Emden, 2013). The way in

Predators and parasitoids of arthropod pests are widely recognized for their role in

78 which these predators interact with, and respond to, pest populations in crop fields varies on a 79 species-specific basis. Particular species of predators may remain closely associated with 80 particular pest populations for extended periods of time (resident predators, e.g. coccinellid and lacewing larvae, anthocorid bugs), or rapidly move among pest populations, and among habitats 81 82 (transient predators, e.g. coccinellid and lacewing adults) (Costamagna and Landis, 2007). 83 Predator identity is an important element of any study that attempts to relate natural enemy 84 activity to pest suppression since intraguild interactions within multi-predator assemblages can vary from positive to neutral to negative, with consequent effects on levels of pest suppression 85 (Cardinale et al., 2003; Ferguson and Stiling, 1996; Henry et al., 2010; Losey and Denno, 1998; 86 Sih et al., 1998; Straub and Snyder, 2006). Consequently, it is difficult to determine species or 87 88 groups of species that are key to suppressing pests within multi-predator assemblages. 89 The soybean aphid, Aphis glycines Matsumura, (Hemiptera: Aphididae), is an introduced pest of soybean that was first detected in North America in 2000 (Ragsdale et al., 2004). 90 91 Soybean aphid has subsequently become a serious economic pest of soybean and has rapidly spread across the soybean producing regions of North America (Ragsdale et al., 2011). In North 92 93 America, the natural enemy guild associated with soybean aphid varies regionally but always 94 contains a significant generalist predator component (Bannerman et al., 2015; Costamagna and 95 Landis, 2007; Desneux et al., 2006; Gardiner et al., 2009b; Hallett et al., 2014; Mignault et al., 96 2006; Ragsdale et al., 2011). More than 50 species of aphidophagous predators and parasitoids 97 feed on A. glycines (Maisonhaute et al., 2017; Ragsdale et al., 2011, and references within), and 98 in many cases provide high levels of suppression (e.g. Fox et al. 2004, 2005, Rutledge et al. 99 2004, Costamagna and Landis 2006, Costamagna et al. 2007). Although the relative abundance and importance of the generalist predators most commonly associated with soybean aphid 100 101 suppression appears to vary regionally, most assemblages studied are numerically dominated by 102 the coccinellids Harmonia axyridis (Pallas), Coccinella septempunctata L., and the anthocorid 103 Orius insidiosus (Say) (Costamagna and Landis, 2007; Desneux et al., 2006; Gardiner et al., 2009b; Hallett et al., 2014; Koch and Costamagna, 2017; Mignault et al., 2006; Ragsdale et al., 104 105 2011). The impact of each predator species within the assemblage on soybean aphid remains unclear, as most previous studies used predator exclusion cages (Costamagna et al., 2007; 106 107 Desneux et al., 2006; Gardiner et al., 2009a), rely solely on correlations (e.g. Maisonhaute et al.,

2017; Yoo and O'Neil, 2009), or used short term assessments of predation (Costamagna and Landis, 2007; Harwood et al., 2009; Petersen and Woltz, 2015; Woltz and Landis, 2013).

The soybean aphid is heteroecious (host alternating), overwintering on woody host plants that are located outside of crop habitats, and migrating into suitable crops in the spring or early summer via the production of alate (winged) morphs (Blackman, 1974; Hodgson et al., 2005b). These colonizing flights are assumed to be relatively local (Schmidt et al., 2012), and soybean aphids continue to produce alate migrants through the summer during all stages of soybean growth and reproduction (Hodgson et al., 2005a, 2005b). The production of alate virginoparae is induced by aphid crowding and declining plant quality (Ríos-Martínez, 2017) and results in both short range dispersal and long range migration. Winged aphids that join previously established colonies contribute directly to population growth rates (Zhang et al., 2009) and large influxes of alatae can disrupt top-down control by natural enemies, increasing aphid densities and thus, population growth rates (Costamagna et al., 2013). However, no studies have examined season-long interactions between alate immigration and predator impact on soybean aphid populations.

Previous studies that examined the impact of natural enemy assemblages on pest populations (see Symondson et al. 2002 for review), including soybean aphids (Costamagna and Landis, 2006; Costamagna et al., 2007; Fox et al., 2005, 2004; Gardiner et al., 2009a; Hallett et al., 2014; Labrie et al., 2016; Maisonhaute et al., 2017; Rutledge et al., 2004; Woltz et al., 2012), were frequently short in duration and often did not distinguish the relative contributions of individual predators within the assemblage. We conducted field population monitoring combined with a manipulative cage experiment for nine weeks (encompassing the period from early aphid colonization to population decline) that allowed us to: 1) identify the key generalist predators associated with soybean aphid throughout the growing season, 2) assess their impact on soybean aphid populations over the season, and 3) determine how summer alate aphid immigration affects season-long aphid population dynamics.

2. Methods

2.1. Sampling locations

The impact of predators and alate aphid immigration on soybean aphid populations was examined at four locations in Minnesota: Rosemount (three commercial soybean fields at the University of Minnesota Outreach Research and Education UMORE Park), LeSeur (three commercial soybean fields), Waseca (two experimental plots at the University of Minnesota Southern Research and Outreach Center and one commercial field), and Saint Paul (three experimental plots at the University of Minnesota St. Paul campus), during 2007. Commercial fields ranged from 14 to 28 ha, and experimental plots ranged from 0.5 to 0.9 ha and their aphid populations were not associated with field size (Linear Regression, $F_{1,10} = 1.65$, P = 0.23). This range of field sizes allows us robust inferences about aphid and predator dynamics and are well within the range of previous studies on this system, that ranged from 0.1 - 0.4 ha plots (Schmidt et al., 2008; Yoo and O'Neil, 2009) to 13.6 – 48.1 ha (Gardiner et al., 2009a). Outbreak populations of soybean aphids resulted in one insecticide application in a commercial field in Waseca (Waseca 3) during the first week of August. Sampling continued in a portion of the field (80 x 330 m) that was left untreated to continue our study. Since predator population trends remained the same in the untreated area and exploratory analysis yielded the same results with or without these data, the analyses presented here include the data from the untreated section of the sprayed field.

2.2. Field observations

Soybean aphid populations were assayed weekly using destructive, whole-plant counts from 12 June to 29 August, with apterous and alate aphids tallied separately. The methods used to count aphids changed over the course of the season in two ways. First, the number of plants per field was reduced (20 plants, 12 June to 13 July; 16, 12 or 8 plants from 16 July to 27 August), a procedure that allowed us to maintain similar levels of precision as plant size and aphid populations increased (Hodgson et al., 2004; Ragsdale et al., 2007). Second, all aphids were counted per plant in June and July, but in August (the last three sampling dates) a node-based sampling method was used to estimate whole-plant aphid abundance (McCornack et al., 2008). Aphid population estimates from this method do not differ from whole-plant counts and consist of weighted estimates based on counts on three plant nodes (all plant material above the newest fully expanded leaf, N₁, the node two node positions below the new growth, N₃, and the middle node of all remaining nodes, N_{MR}).

Predators were sampled weekly from 12 June until 27 August during a two-hour period between 8:30 and 16:30. Three methods were used to sample predators and parasitoid mummies: sweep-netting, destructive whole-plant counts, and 25 m walking transects (Bannerman et al., 2015). Each sweep-net sample consisted of 25, 180° sweeps through the soybean canopy along a linear transect, with 4 samples taken per field per week. Destructive whole-plant counts consisted of visual inspection, both before disturbance and after uprooting, of each plant on which aphid counts occurred. Each walking transect consisted of an observer walking a 25-m transect along linear soybean rows and recording all observed predators and parasitoid mummies that were visible on the upper leaves of each plant (736 ± 11 (mean \pm SD) soybean plants per 25m of row). The number of transect samples mirrored the number of wholeplant samples as the two were performed simultaneously. Transect counts were considered superior for sampling large predators like coccinellids, and destructive whole-plant counts superior for sampling small predators like O. insidiosus (Bannerman et al., 2015). Inclement weather prevented field level sampling on two occasions, from 9-13 July at Rosemount, and from 30 July to 3 August at St. Paul. The predator abundance data presented here was previously published in a study that compared sampling methods for soybean aphid predators (Bannerman et al., 2015). The presence of entomopathogenic fungi was not assessed, but no evidence of fungal epizootics was observed in any field.

2.3. Exclusion study

To determine the impact of predators on soybean aphid populations, exclusion cages were used to protect aphids from predation. Pairs of field plants were selected bearing numbers of aphids that approximated the mean aphid density in the field (based on the destructive plant counts). Each plant was then randomly assigned to one of two treatments: open plant or predator exclusion cage. Three sets of plants were established in each field every sampling week. In some cases, aphids were physically removed to better approximate the target aphid density. Open plants consisted of a marked soybean plant with the surrounding plants removed to eliminate plant-to-plant contact and provide more similar growing conditions as plants with exclusion cages. Previous studies using sham cages have demonstrated that removal of the surrounding plants around open plants, which may reduce between-plant movement of immature predators, has no meaningful effect on aphid densities (Costamagna et al., 2008). Exclusion

cages were constructed with cylindrical tomato wire cages (0.4 x 1.0 m), secured to the ground with four commercial plastic tent pegs passing through wire loops made at the base of the cage and covered with white no-see-um mesh (Kaplan Simon Co., Baintree, MA) (after Costamagna and Landis 2011). The cages were further secured by tying them to two sturdy stakes (1.52 m) located on opposite sides. Previous work indicated that this type of cage does not alter microclimatic conditions sufficient to change interactions among plants and aphids (Costamagna et al., 2013).

Apterous (wingless) and alate (winged) morphs were both counted one week after the imposition of treatments. Since new plants were assigned to treatments every week, the impact of predators could be partitioned weekly over the duration of the study. This procedure also reduced variance associated with aphid overcrowding and declining plant quality, which tends to occur when natural enemies are excluded from aphid colonies for long periods (Kucharik et al., 2016).

The start date of the exclusion study differed among locations, commencing during the week of 11-15 June in the three fields at St. Paul, and between 18-22 June in all other fields. Exclusion studies ended during the week of 23-27 July in St. Paul fields, and the week of 6-10 August in all other fields, resulting in three fields with seven weeks of cage manipulations and nine fields with nine weeks. Because the duration of the cage experiments (7-9 weeks) was shorter than the duration of field-level assessments (11 weeks), any mention of 'week' in this paper refers to field-level assessment dates.

Since we initiated new cages each week for the duration of this experiment it was important that initial aphid density on caged plants was representative of the field mean. We were successful in achieving this objective with one caveat. In week three, the number of aphids on caged plants averaged 3.5 aphids (range = 3 to 5), higher than that of the field mean in the LeSeur and Waseca fields. This was necessary because aphid densities in these fields averaged < 1 aphid per plant, and caging plants with only one or two aphids would provide no useful information. Overall, these slightly inflated initial values appear to have little impact on conclusions derived from our models (based on running models that include and exclude data from these early weeks) since they occurred when both aphid and predator populations were very low. For simplicity, we only show results including these fields.

There was a secondary influx of alate aphids into all fields in mid- to late-July which increased alate abundance. Furthermore, clearing immediately surrounding 'open' plants resulted in higher than normal attraction of alates, resulting in alatae densities on open plants well above the mean field density. This was probably a consequence of 'edge' or 'bare ground' effects, which are often observed in other aphid species (Davis et al., 2015, 2009; Irwin et al., 2007). This immigration event interacted with our open plant treatment, providing us with an opportunity to compare predation under 'normal' immigration levels (i.e. aphid counts on unmanipulated plants, hereafter referred to as field counts) with predation under high immigration levels (i.e. aphid counts in open plant treatment, hereafter referred to as open plant counts).

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2.4. Analysis

Generalized linear models (GLMs) were constructed to assess the effects of predators and alate aphid immigration (dependent variables) on apterous aphid density (independent variables) through the growing season. We compared the fit of negative binomial and poisson distributions, both theoretically appropriate for count data, using the deviance/df ratio and diagnostic graphs. The negative binomial distribution fit best in all cases, so we used it for all models presented here. Several sets of predator predictor variables were tested: (1) A combination of destructive whole-plant counts for small predators and 25 m walking transect counts for large predators, (2) Sweep net counts for all predators, (3) Natural enemy units (a method that normalizes the impact of soybean aphid natural enemies into a single value), each based on recent soybean aphid natural enemy sampling literature (Table 1) (Bannerman et al., 2015; Hallett et al., 2014). Additional control predictor variables in all models included aphid density in the exclusion cage (to control for variable initial aphid numbers and environmental variables that may affect aphid population growth), field, and week. Initial exploratory models involved mixed models (GLMMs), and included 'field' nested within 'site', treated as a random variable. However, this did not improve model fit (using AIC as the index), so we did not include it in our analyses. Furthermore, including various combinations of random variables in models (i.e., 'field' as a random variable excluding 'site', and 'field' as a fixed variable), did not change any conclusions, so we decided to present the simplest models based on AIC. All

analyses were performed using the Mass package in R version 3.2.1 (R Core Team, 2015; Venables and Ripley, 2002).

Although we constructed exploratory models to test for the impact of most predatory taxa observed, the models presented here focus on only three species: *H. axyridis*, *C. septempunctata*, and *O. insidiosus*. These three species represented 65% of the total predators observed using either sweep-netting or a combination of destructive whole-plant counts and 25 m walking transects. These three predators have been reported as important natural enemies of soybean aphid across North America (Costamagna and Landis, 2007; Desneux et al., 2006; Gardiner et al., 2009b; Hallett et al., 2014; Koch and Costamagna, 2017; Mignault et al., 2006; Ragsdale et al., 2011). In addition, we observed too few individuals of many species to reasonably include them as an independent variable. We also constructed exploratory models that included larval *H. axyridis* and *C. septempunctata* counts as independent variables and as combined variables (as done with *O. insidiosus*). Larval coccinellid counts were not significant predictors of apterous aphid abundance and since the inclusion of these predictors did not change the significance or direction of response of any other predictor variable, the models are not shown or discussed further.

Since alate aphid immigration influenced aphid density in the open plant treatments, we constructed two sets of models: the first set used field-based aphid counts for apterous and alate aphid densities (field models, normal immigration levels) and the second set used open plant-based aphid counts (open models, high immigration levels). We then used the parameter estimates from two models to predict aphid abundance in response to a range of *H. axyridis* abundances to illustrate the strength of the predator-prey interaction in this system. This required a two-step process. First, to model *H. axyridis* abundance, we constructed a set of representative numerical responses (zero, low, medium and high), approximating the range of responses observed across our 12 focal fields. We then predicted the effect of *H. axyridis* abundance on apterous aphid abundance while setting all other predictor variables to their mean weekly values (i.e. we varied the intensity of the *H. axyridis* numerical response while keeping the values of all other predictor variables constant to isolate and illustrate the impact of *H. axyridis* on apterous aphid abundance). Since 'field' was also a significant predictor of apterous aphid abundance, we show the results for three fields representative of low, intermediate and high peak aphid densities, respectively.

3. Results

3.1. Soybean aphids

Although all 12 fields had been colonized by soybean aphids by 15 June (week 1), peak aphid abundance and its timing varied (Figure 1). Peak abundance ranged from 144.8 ± 95.8 to 1726.2 ± 897.1 aphids per plant (mean \pm SEM), providing us a broad range of aphid population levels to investigate relationships with predator assemblages. For all fields and sample dates, alate aphids represented less than 5% (mean = 0.77%, SD = 0.11%) of the aphid population at field level, but up to 23% (mean = 2.3%, SD = 3.1%) on the open plants. Peak aphid abundance occurred between 23 July and 15 August (weeks 7, 8, 9 & 10) while soybean plants were in the R3, R4 or R5 reproductive stage, and was followed by swift declines in all fields by the end of August. Peak aphid abundance occurred during the cage experiment for 11 of 12 study fields (Figure 1).

3.2. Predator responses

The most abundant predators collected throughout the season were the coccinellids *H. axyridis* and *C. septempunctata* and the anthocorid bug *O. insidiosus*, collectively representing 65% of the aphidophagous predators captured using sweep-nets or observed with a combination of 25 m walking transects and destructive whole-plant counts across all fields and sampling dates (Bannerman et al., 2015). The abundances of these predators changed throughout the season and varied among fields (Figure 2, and Appendix A Figures A.1 and A.2), but reflected the range of abundances previously reported from soybean in the Midwest USA (see references in Bannerman et al. 2015). Peak *H. axyridis* densities varied among fields, ranging from 0.25 to more than 30 adult beetles per transect. We observed a distinct numerical response by *H. axyridis* in 11 of 12 fields with peak abundance occurring between 0 and 3 weeks after peak aphid abundance. Synchronized numerical responses of *C. septempunctata* and *O. insidiosus* to increasing aphid abundance were observed in some fields but were inconsistent (Figures A.1 and A.2). Though plots of predators abundances based on sweep-netting are not provided, the patterns described are similar to those reported based on walking transects. There were no parasitoid mummies observed in any field in Minnesota in 2007.

3.3. Exclusion study

All models indicated a non-linear increase in aphid populations followed by a decrease with significant 'week' and 'week²' terms, and various levels of aphids with significant field terms (Table 2 and Appendix A Tables A.1 and A.2). In models that measured predators using a combination of transect counts and whole-plant counts, apterous aphid abundance was negatively associated with adult *H. axyridis* abundance in both unmanipulated field plants (Estimate = -0.065, SE = 0.024, p = 0.006), and open plants (Estimate = -0.076, SE = 0.021, p = 0.0002; Table 2). No relationships were observed between apterous aphid abundance and adult *C. septempunctata* or adult + juvenile *O. insidiosus* abundance. Numbers of aphids in exclusion cages and numbers of alate aphids in fields did not predict apterous aphid abundance significantly in our field-based model, but did in our open model (aphid counts in exclusion cages: Estimate = 0.0008, SE = 0.0002, p < 0.0001; alate aphid counts on open plants: Estimate = 0.042, SE = 0.011, p < 0.0001). This demonstrated the impact of higher than normal alate aphid abundance on our 'open cage' plants. In both models, adult *H. axyridis* abundance was the only predictor to be negatively associated with soybean aphid abundance.

Additional models were constructed to determine whether the above results were robust or dependent on the specific predator/sampling method variables used. Field and open models using sweep-count data for all three predatory species produced similar models to those using transect and plant count data, with one difference. In the field + sweep-net model, apterous aphid abundance was positively associated with *O. insidiosus* abundance, supporting other models in our study that suggest this predator does not negatively affect aphid population growth, although it does exhibit a functional response to aphid density (Appendix A: Table A.1). The natural enemy units predictor variable did not significantly affect apterous aphid populations in either the field or open models (Appendix A: Table A.2). In summary, all models constructed and variables tested suggest that the only predator with consistent negative associations with aphid abundance was adult *H. axyridis*.

Modeling aphid population responses to *H. axyridis* revealed the strength of the impact of this predator in the system. The abundance of *H. axyridis* observed across the 12 sample fields suggested that numerical responses peaking at 1, 5, and 25 adults / transect would be realistic representations of the range of responses in our area of study (Figures 2 and 3). We used a null response (i.e., zero *H. axyridis*) and these three numerical responses to predict aphid populations

using mean weekly values and model parameters for three fields representing a range of aphid populations. Both field and open models predicted that a high numerical response of *H. axyridis* will result in a strong aphid population reduction in all three cases (Figure 4). Intermediate and low numerical responses by *H. axyridis* resulted in either slowed aphid population growth or reduced aphid populations when compared with models without *H. axyridis*, although to a lesser extent than observed with a high numerical response. Under normal alate immigration levels (field models), seven modelled populations with *H. axyridis* reached the economic threshold (ET) of 250 aphids per plant, but only three exceeded the economic injury level (EIL) of 674 aphids per plant (Ragsdale et al., 2007) (Figures 4a-c). Similarly, eight modelled populations exceeded the ET and five populations exceeded the EIL in models under high alate immigration (open models, Figures 4d-f). High alate aphid immigration levels resulted in an accelerated growth and decline of aphid populations (Figures 4d-f).

4. Discussion

Our study demonstrated that generalist predators can exert season-long suppression of aphids, even under high rates of alate immigration. We found that within an assemblage dominated by three generalist predators, H. axyridis was the critical species, exhibiting consistent, season-long negative associations with soybean aphid populations in Minnesota. Throughout the season, generalist predators from 12 insect and 2 arachnid families were recorded in Minnesota soybean fields with the coccinellids H. axyridis and C. septempunctata and the anthocorid bug O. insidiosus being most abundant. These three species have been identified in previous studies as potentially important soybean aphid predators in several regions in North America (Costamagna and Landis, 2007, 2006; Fox et al., 2005, 2004; Koch and Costamagna, 2017; Labrie et al., 2016; Maisonhaute et al., 2017; Rutledge and O'Neil, 2005; Rutledge et al., 2004). Though these three species were all abundant, our results confirm previous studies suggesting that coccinellids were the key predators suppressing A. glycines (Costamagna and Landis, 2007; Costamagna et al., 2008, 2007), although the present results suggest a key role of *H. axyridis*, but not *C. septempunctata*, in contrast with previous observations in this system (Costamagna and Landis, 2007). This discrepancy may result from the use of smaller spatial or temporal scales in previous studies of aphid suppression by C.

septempunctata, such as individual or groups of fireweed stems (Ives et al., 1993) or 1-m² soybean patches (Costamagna and Landis, 2007), and typically for observation periods of a few hours or days. Our results suggest that at the whole-field level over season-long scales, *C. septempunctata* does not show an association with *A. glycines* suppression.

Similarly, previous studies suggesting that *O. insidiosus* can suppress soybean aphid populations focused on their impact on early-season aphid populations, rather than on season-long dynamics (Brosius et al., 2007; Yoo and O'Neil, 2009). Although we did not consistently detect statistically significant patterns of association between *O. insidiosus* and aphid abundance, their abundance did increase in response to increasing aphid populations in most fields, suggesting that larger aphid populations will attract larger numbers of this predator. The results demonstrate that *H. axyridis* can contribute to both prevention and suppression of *A. glycines* outbreaks throughout the growing season (Koch and Costamagna, 2017), and underscore the importance of studies that can identify and quantify the impact of particular species within a natural enemy assemblage (Symondson et al., 2002) since presence alone, even in high abundance, does not necessarily imply pest population reduction.

In order for generalist predators to significantly reduce pest populations, theory suggests that either early arrival (while pest populations are still small) or a strong numerical response is key (Bianchi et al., 2006; Chang and Kareiva, 1999; Rutledge et al., 2004). For H. axyridis feeding on soybean aphid, a strong numerical response occurred in most fields near the end of July or early August, between 0 and 2 weeks after peak aphid abundance. Peak H. axyridis abundance was higher in fields with higher peak aphid populations, further demonstrating that adult *H. axyridis* are attracted to higher aphid densities. The densities of *H. axyridis* observed in our study were also well within the range observed in previous studies (e.g. Costamagna and Landis 2006, Desneux et al. 2006). Given variation in the strength of the H. axyridis numerical response, we modelled four response levels, all within the observed range of responses, to demonstrate the potential impact of *H. axyridis* on aphid populations. The results of this exercise demonstrated that H. axyridis can control aphids at low/intermediate aphid infestation levels, but that high aphid populations can still escape control if *H. axyridis* fails to respond in sufficiently high numbers. For perspective, an increase of one H. axyridis adult per transect count is approximately equivalent to one 'visible' beetle (i.e. visual samples on transects only reflect adult beetles on the upper side of upper leaves, see Bannerman et al. 2015 for more details) per 736 \pm

11 soybean plants (mean number of soybean plants per 25 m of one row of soybean), so although the *H. axyridis* parameter estimates are small in our models, the per capita effect size of *H. axyridis* on aphid populations is large, even at densities well below 1 visible adult per 30 plants.

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Secondary influxes of alate aphids have been associated with outbreak populations in soybean aphid (Hodgson et al., 2005b; Labrie et al., 2016; Rhainds et al., 2010). Surprisingly, only when artificially elevated rates of alate immigration occurred (on open treatment plants) did this immigration increase apterous aphid populations, and even then, it did not counter the impact of *H. axyridis* on aphid numbers. Thus, our results suggest that aphid populations can still be suppressed by predators despite continued alate immigration.

Our study supports the view that naturally occurring generalist predators can significantly reduce agricultural pest populations (Murdoch, 1994; Symondson et al., 2002) and has several implications for agricultural pest management. First, even though all twelve fields sampled experienced a period of rapid aphid population growth, this did not guarantee that aphid populations would exceed the economic injury level (Ragsdale et al., 2007), which is at least partially attributable to the activity of generalist predators. Second, as efforts advance to integrate measures of natural enemy abundance into economic decision making (e.g. Elliott et al. 2004, Giles and Walker 2009, Hallett et al. 2014), it will become increasingly important to predict how natural enemy abundance affects pest populations in time and space. We did not detect a relationship between natural enemy units (as per Hallett et al. 2014) and apterous aphid abundance. This was likely caused by combining the impact (abundance x voracity) of one species with a strong numerical response to aphids with that of two species that did not, thereby weakening our ability to relate natural enemy abundance to aphid abundance. In addition, during our 2007 studies we observed no parasitoids and these comprised a larger component of the natural enemy assemblage in Ontario where the natural enemy unit concept was developed and validated (Hallett et al., 2014), and parasitoids of soybean aphids are now increasing in other regions in North America (Kaser, 2016). Therefore, regional differences in the assemblage of soybean aphid natural enemies may affect the applicability of methodology developed to assess their impact in one particular region. Similarly, temporal changes in natural enemy assemblages, particularly an increase in parasitism by the adventive parasitoid *Aphelinus certus* (Hymenoptera: Aphelinidae), should be addressed in future studies (Kaser, 2016).

Our study demonstrated the important role of *H. axyridis*, a transient generalist predator, in reducing soybean aphid populations, adding to evidence that naturally occurring generalist predators can regulate aphid populations, even under levels of high alate immigration. We also showed that the numerical response of *H. axyridis* is strong when aphid abundance is high. At the same time, we found that some generalist predators, even though abundant, may have little impact on increasing aphid populations. These findings should ultimately be useful to improve decision-making in pest management programs as we seek to enhance natural pest control services through the development and implementation of dynamic action thresholds (e.g. Hallett et al. 2014) and conservation biocontrol (Landis et al., 2000).

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464	Author contributions
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466	ACC, BPM and DWR designed the experiments; ACC and BPM performed the experiments; JB,
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Table Captions

- Table 1. Predator predictor variable sets used for GLM models to examine the impacts of
- predators on season-long apterous aphid populations in 12 Minnesota soybean fields from 25
- 670 June 2007 10 August 2007.
- Table 2. Relationship between predator and aphid populations using transect + plant count
- 672 predator predictor variables for 12 Minnesota soybean fields from 25 June 2007 10 August
- 673 2007.

Tables

675 Table 1.

Variable description	Justification		
Mean # of adult H. axyridis per transect	Combination of the two methods determined to be		
Mean # of adult C. septempunctata per transect	efficient and effective for sampling both large and small predators of soybean aphid (Bannerman et al.		
Mean # of juvenile + adult O. insidiosus per plant	2015)		
Mean # of adult H. axyridis per 25 sweeps	Sweep netting represents a good method for		
Mean # of adult C. septempunctata per 25 sweeps	sampling both large and small natural enemies of soybean aphid when efficiency is not paramount		
Mean # of juvenile + adult O. insidiosus per 25 sweeps	(Bannerman et al. 2015)		
	Method developed by Hallett et al. (2014) to normalize the impact of soybean aphid natural enemies into a single value. 1 NEU represents 100 aphids consumed or parasitized per day and is calculated using the equation:		
Mean natural enemy units per plant	$NEU_{total} = \sum_{i=1}^{N} n_i V_i$		
	Where $_{ni}$ is the total number of individuals of species i and V_i is the mean voracity of species i divided by 100. Our NEU calculations used the mean abundance of juvenile and adults stages of H . axyridis, C . septempunctata and O . insidiosus per destructive plant count.		

678 Table 2.

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Apterous aphid = Aphid density exclusion + Alate density + H. axyridis adult + C. septempunctata adult + O. insidiosus all stages + Week + Week² + Field^a Transect + plant count Field counts^b Z-value Estimate Standard error *p*-value 2.33 x 10⁻⁶ Intercept -3.448 0.730 -4.72 7.69 x 10⁻² Aphid exclusion cage 0.0003 0.0002 1.77 Alate field 0.0777 0.0913 0.85 0.39 -2.74 6.06×10^{-3} H. axyridis adult transect -0.0650 0.024 C. septempunctata adult transect -0.0953 -0.74 0.46 0.129 O. insidiosus all stages plant count -0.340 0.211 -1.61 0.11 $< 2.0 \times 10^{-16}$ Week 0.253 8.31 2.102 Week² 4.06 x 10⁻⁷ -5.07 -0.104 0.021 Open plant counts^c Standard error Estimate Z-value p-value Intercept -0.599 0.558 -1.070.28 1.52 x 10⁻⁶ Aphid exclusion cage 0.0008 0.0002 -0.57 9.04 x 10⁻⁵ Alate open plant 0.0419 0.0107 3.92 2.09 x 10⁻⁴ H. axyridis adult transect -0.0762 0.0206 -3.71 C. septempunctata adult transect -1.49 0.14 -0.169 0.114 O. insidiosus all stages plant count -0.101 0.178 -0.57 0.57 2.88 x 10⁻¹¹ Week 1.334 0.201 6.65 Week² 1.74 x 10⁻⁵ -0.0718 0.0167 -4.30

^a For simplicity, estimates for the effect of each field on aphid abundance not shown. Field is a significant predictor of Apterous aphid abundance (Analysis of deviance: Field counts model, df = 11, p < 0.0001; Open plant counts model, df = 11, p < 0.0001)

^b Apterous aphid response variable and Alate aphid predictor variable based on field counts

^c Apterous aphid response variable and Alate aphid predictor variable based on open plant counts

Figure Captions

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Figure 3 for *H. axyridis* numerical responses.

681 Figure 1. Mean (± SEM) weekly soybean aphid abundance per plant observed in 12 Minnesota soybean fields from 25 June 2007 – 10 August 2007. Line below x-axis indicates duration of 682 683 cage experiment in the respective field. 684 Figure 2. Mean (± SEM) weekly adult *Harmonia axyridis* abundance per 25 m walking transect observed in 12 Minnesota soybean fields from 25 June 2007 – 10 August 2007. 685 686 Figure 3. Modeled numerical response intensities for *Harmonia axyridis*. Value ranges were set 687 to represent the range of responses observed in Minnesota soybean fields in 2007 and used for 688 predictive purposes to visualize the effect of H. axyridis on soybean aphids (Figure 4) based on 689 our GLM models. Each transect represents 25 linear-m of soybean. Figure 4. Predicted effect of *Harmonia axyridis* on apterous aphid abundance for three 690 691 representative soybean fields using Field (subplots A,B,C) and Open models (subplots D,E,F) 692 (Table 2). The low aphid population field was Waseca 2 (peak density close to the ET); the 693 intermediate aphid population field was Rosemount 3 (peak aphid density > ET, < EIL); and the 694 high aphid population field was Rosemount 1 (peak density >> EIL). Dotted line = No numerical 695 response by *H. axyridis*; Dot-dash line = low intensity numerical response; dashed line = 696 intermediate intensity numerical response; solid line = high intensity numerical response (see

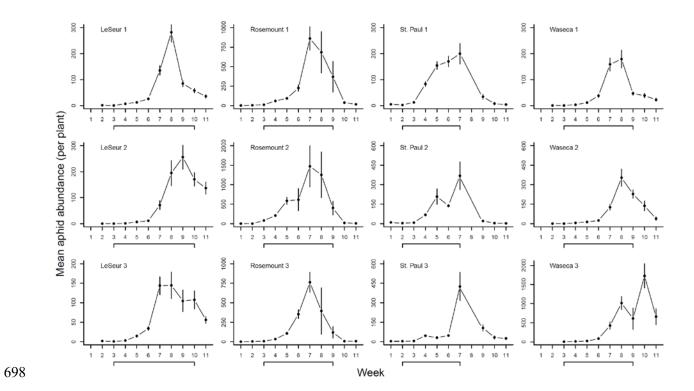


Figure 1. Mean (\pm SEM) weekly soybean aphid abundance per plant observed in 12 Minnesota soybean fields from 25 June 2007 – 10 August 2007. Line below x-axis indicates duration of cage experiment in the respective field.

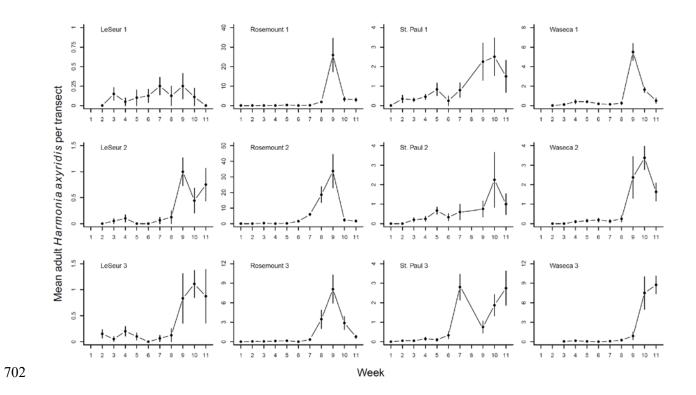


Figure 2. Mean (\pm SEM) weekly adult *Harmonia axyridis* abundance per 25 m walking transect observed in 12 Minnesota soybean fields from 25 June 2007 – 10 August 2007.

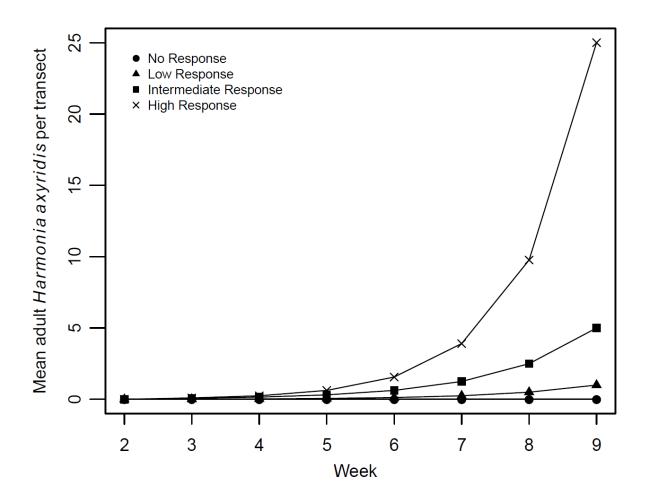


Figure 3. Modeled numerical response intensities for *Harmonia axyridis*. Value ranges were set to represent the range of responses observed in Minnesota soybean fields in 2007 and used for predictive purposes to visualize the effect of *H. axyridis* on soybean aphids (Figure 4) based on our GLM models. Each transect represents 25 linear-m of soybean.

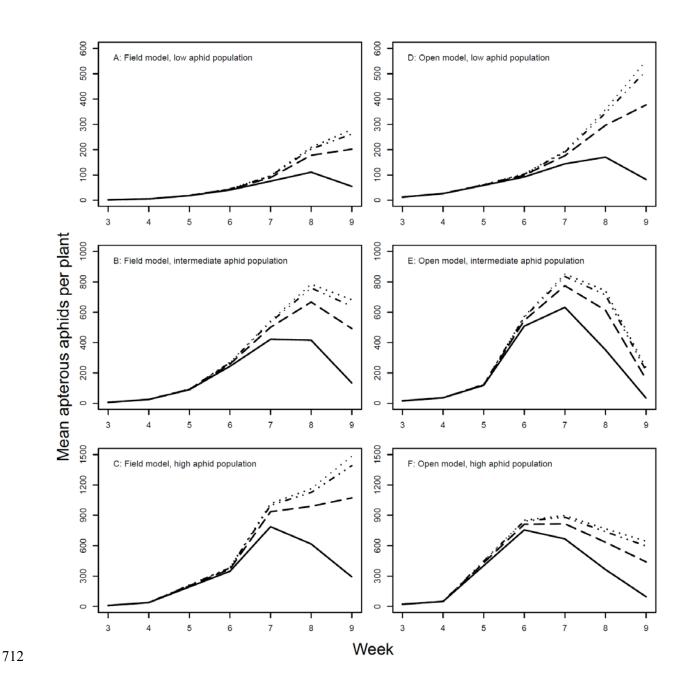


Figure 4. Predicted effect of *Harmonia axyridis* on apterous aphid abundance for three representative soybean fields using Field (subplots A,B,C) and Open models (subplots D,E,F) (Table 2). The low aphid population field was Waseca 2 (peak density close to the ET); the intermediate aphid population field was Rosemount 3 (peak aphid density > ET, < EIL); and the high aphid population field was Rosemount 1 (peak density >> EIL). Dotted line = No numerical response by *H. axyridis*; Dot-dash line = low intensity numerical response; dashed line = intermediate intensity numerical response; solid line = high intensity numerical response (see Figure 3 for *H. axyridis* numerical responses.

Appendix A

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Table A.1. Relationship between natural enemy and aphid populations using sweep-net count natural enemy predictor variables for 12 Minnesota soybean fields from 25 June 2007 - 10 August 2007.

Apterous aphid = Aphid density exclusion + Alate density + <i>H. axyridis</i> adult + <i>C. septempunctata</i> adult + <i>O. insidiosus</i> all stages + Week + Week ² + Field ^a							
Sweep-net							
Field counts ^b	Estimate	Standard error	Z-value	<i>p</i> -value			
Intercept	-2.2541	0.754	-2.99	2.78 x 10 ⁻³			
Aphid exclusion cage	-0.0001	0.0002	-0.41	0.68			
Alate field	-0.0242	0.0761	-0.32	0.75			
H. axyridis adult sweep-net	-0.1864	0.0372	-5.01	5.55 x 10 ⁻⁷			
C. septempunctata adult sweep-net	0.1706	0.197	0.86	0.39			
O. insidiosus all stages sweep-net	0.1820	0.0448	4.07	4.81 x 10 ⁻⁵			
Week	1.6700	0.251	6.66	2.69 x 10 ⁻¹¹			
Week ²	-0.0661	0.0211	-3.13	1.76 x 10 ⁻³			
Open plant counts ^c	Estimate	Standard error	<i>Z</i> -value	<i>p</i> -value			
Intercept	0.1733	0.641	0.27	0.79			
Aphid exclusion cage	0.0006	0.0002	3.32	9.08 x 10 ⁻⁴			
Alate open plant	0.0441	0.0109	4.04	5.25 x 10 ⁻⁵			
H. axyridis adult sweep-net	-0.1280	0.0373	-3.43	6.03 x 10 ⁻⁴			
C. septempunctata adult sweep-net	-0.0087	0.186	-0.05	0.96			
O. insidiosus all stages sweep-net	0.0978	0.0450	2.17	0.03			
Week	1.0503	0.216	4.88	1.09 x 10 ⁻⁶			
Week ²	-0.0479	0.0190	-2.52	0.01			

^a For simplicity, estimates for the effect of each field on aphid abundance not shown. Field is a significant predictor of Apterous aphid abundance (Analysis of deviance: Field counts model, df = 11, p < 0.0001; Open plant counts model, df = 11, p < 0.0001)

^b Apterous aphid response variable and Alate aphid predictor variable based on field counts

^c Apterous aphid response variable and Alate aphid predictor variable based on open plant counts

Table A.2. Relationship between natural enemy and aphid populations using NEU plant count natural enemy predictor variables for 12 Minnesota soybean fields from 25 June 2007 – 10 August 2007.

Apterous aphid = Aphid density exclusion + Alate density + NEU + Week + Week ² + Field ^a							
Natural enemy units (per plant)							
Field counts ^b	Estimate	Standard error	<i>Z</i> -value	<i>p</i> -value			
Intercept	-3.063	0.737	-4.16	3.20 x 10 ⁻⁵			
Aphid exclusion cage	0.0004	0.0002	1.92	5.49 x 10 ⁻²			
Alate field	0.140	0.0853	1.64	0.10			
NEU	-0.125	0.536	-0.23	0.82			
Week	1.987	0.255	7.78	7.14 x 10 ⁻¹⁵			
Week ²	-0.106	0.0210	-5.07	4.06 x 10 ⁻⁷			
Open plant counts ^c	Estimate	Standard error	<i>Z</i> -value	<i>p</i> -value			
Intercept	-0.538	0.569	-0.94	0.35			
Aphid exclusion cage	0.0009	0.0002	4.86	1.18 x 10 ⁻⁶			
Alate open plant	0.0543	0.0111	4.91	9.10×10^{-7}			
NEU	0.254	0.488	0.52	0.60			
Week	1.289	0.204	6.34	2.38 x 10 ⁻¹⁰			
Week ²	-0.0773	0.0171	-4.53	5.84 x 10 ⁻⁶			

^a For simplicity, estimates for the effect of each field on aphid abundance not shown. Field is a significant predictor of Apterous aphid abundance (Analysis of deviance: Field counts model, df = 11, p < 0.0001; Open plant counts model, df = 11, p < 0.0001)

^b Apterous aphid response variable and Alate aphid predictor variable based on field counts

^c Apterous aphid response variable and Alate aphid predictor variable based on open plant counts

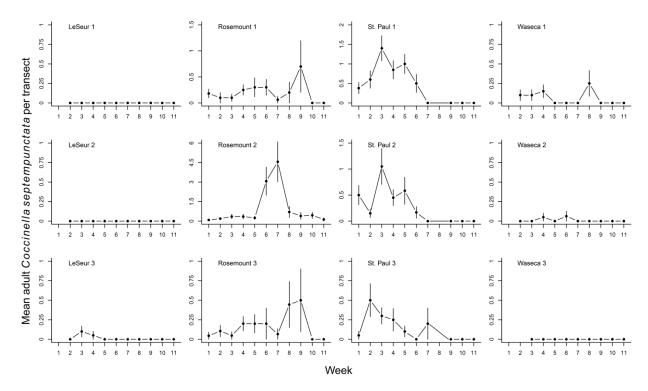


Figure A.1. Mean (± SEM) weekly adult *Coccinella septempunctata* abundance per transect observed in 12 Minnesota soybean fields from 25 June 2007 – 10 August 2007.

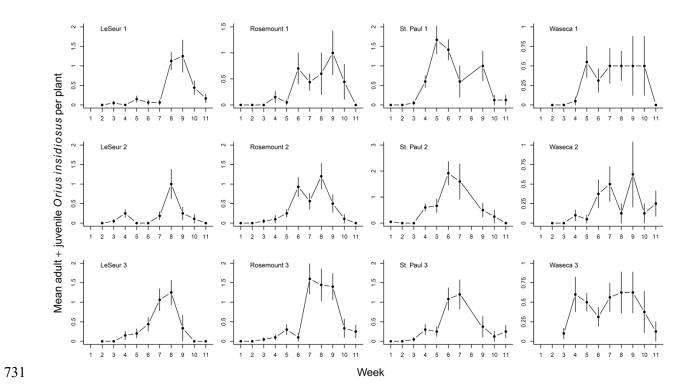


Figure A.2. Mean (± SEM) weekly adult + juvenile *Orius insidiosus* abundance per plant observed in 12 Minnesota soybean fields from 25 June 2007 – 10 August 2007.